

ECOLE D'ETE GN-MEBA

Microscopie Electronique à Balayage et Microanalyses

Analyse d'images, traitement des données 3D

Marco Cantoni

EPFL-CIME

Organisation :



GEOSYSTEMES

Supports techniques :



SYNERGIE⁴



Autres supports :



outline

- Why image processing, and how...?
- Image processing in 2D
 - What is an ideal image ...? Histogram tells stories...!
 - Before taking the image: the right imaging conditions !
- Tools for 2D image analysis/processing
- 3D volume reconstruction by FIB/SEM
- Processing steps for 3D reconstruction
- Registration, Segmentation, Quantification
- Visualisation

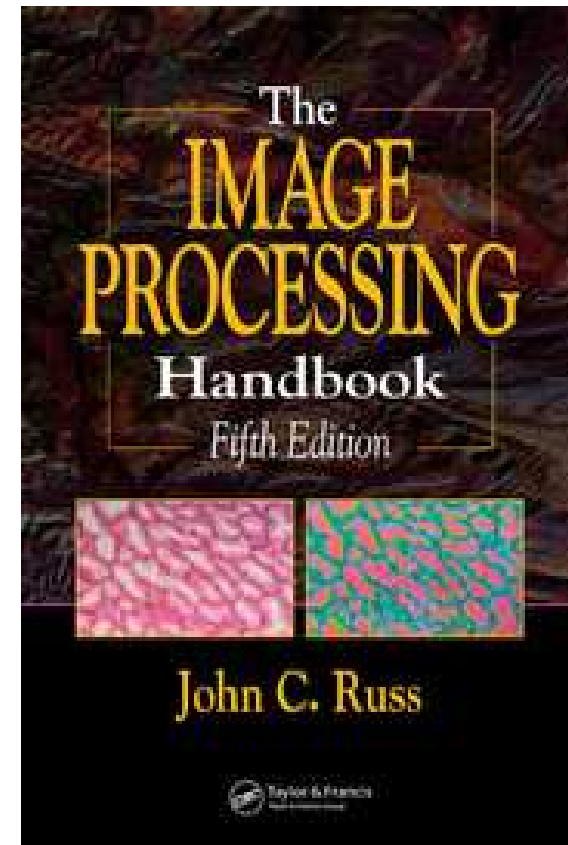
Image processing basics

“the bible”

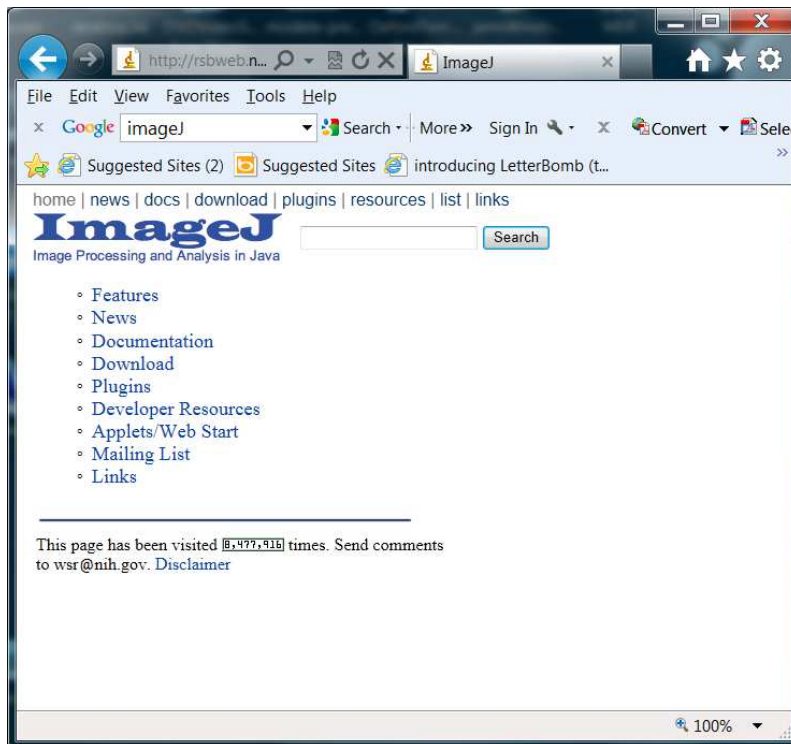
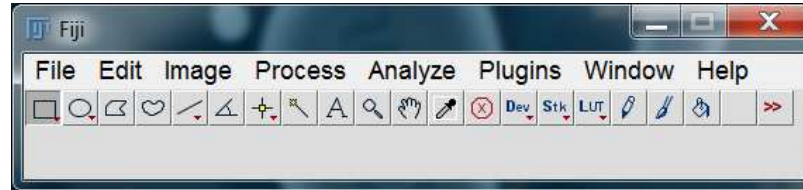
John C. Russ (2002)

*The Image Processing
Handbook*

5th edition, CRC Press



The (free) tools

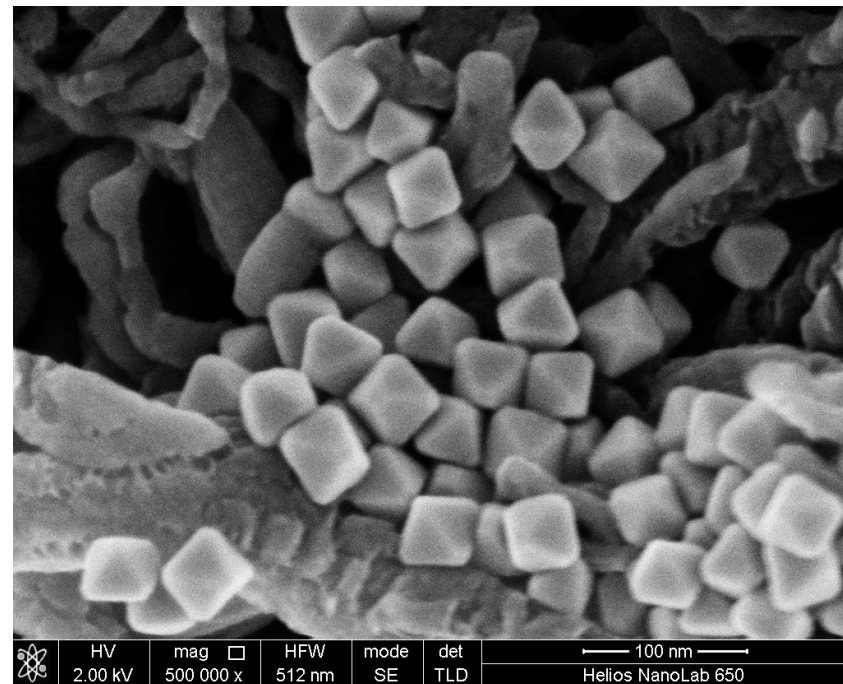
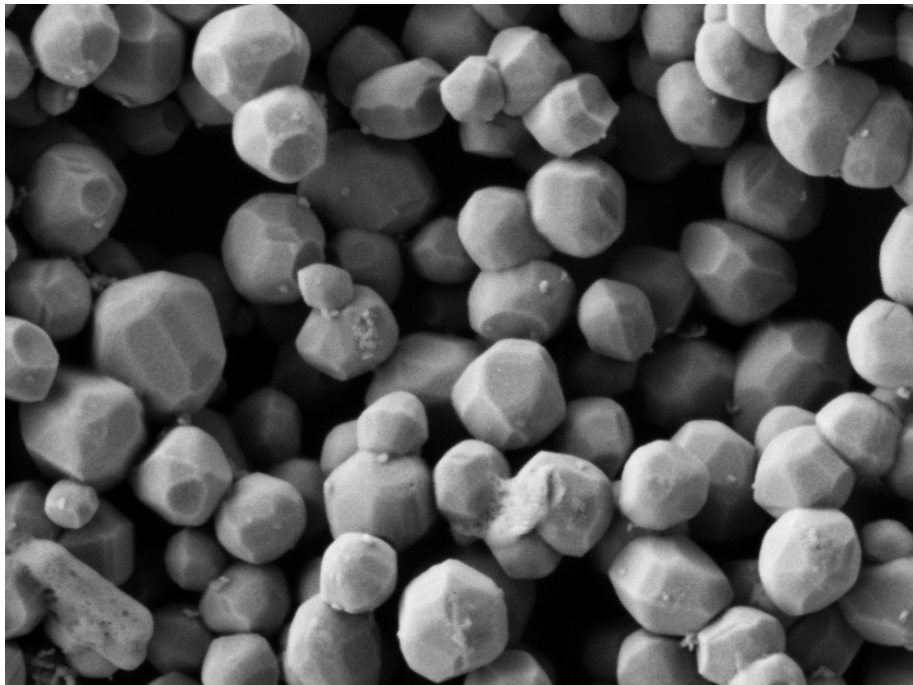


<http://rsbweb.nih.gov/ij/>

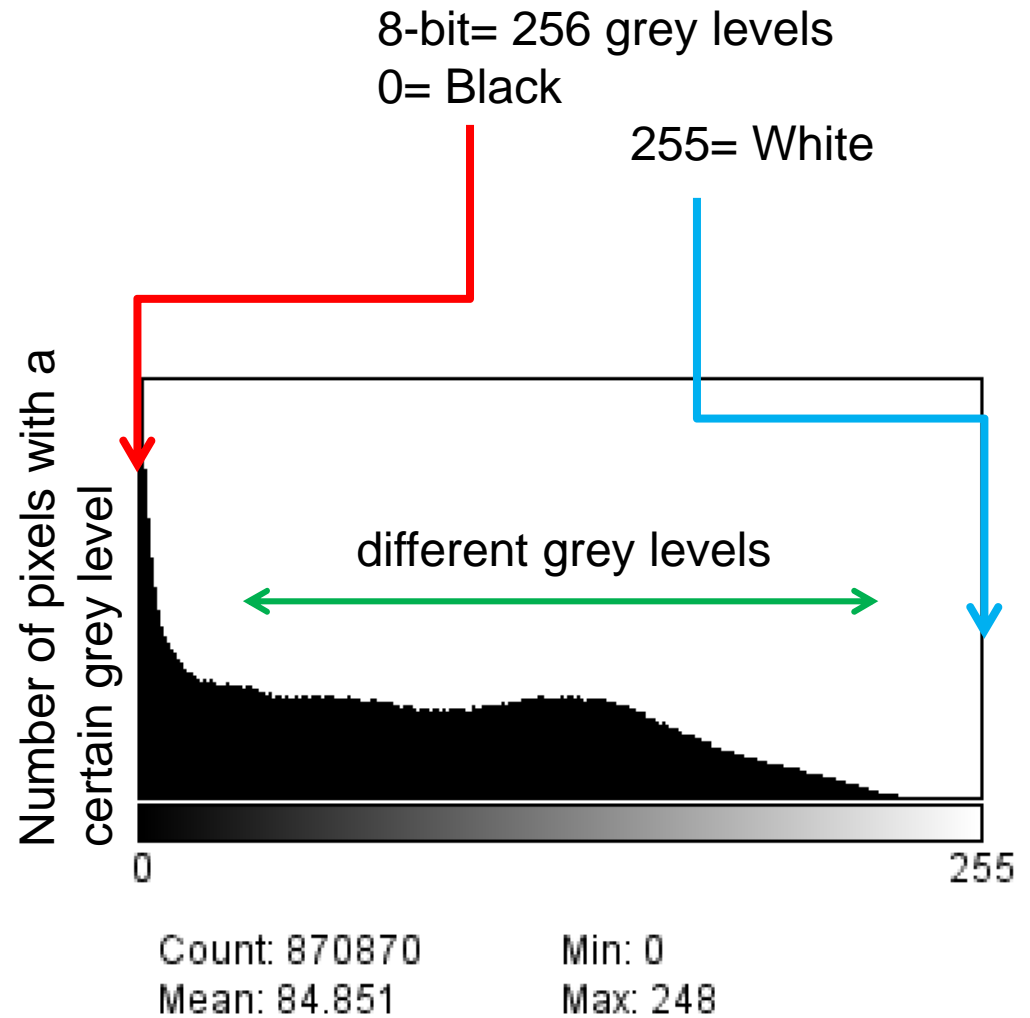
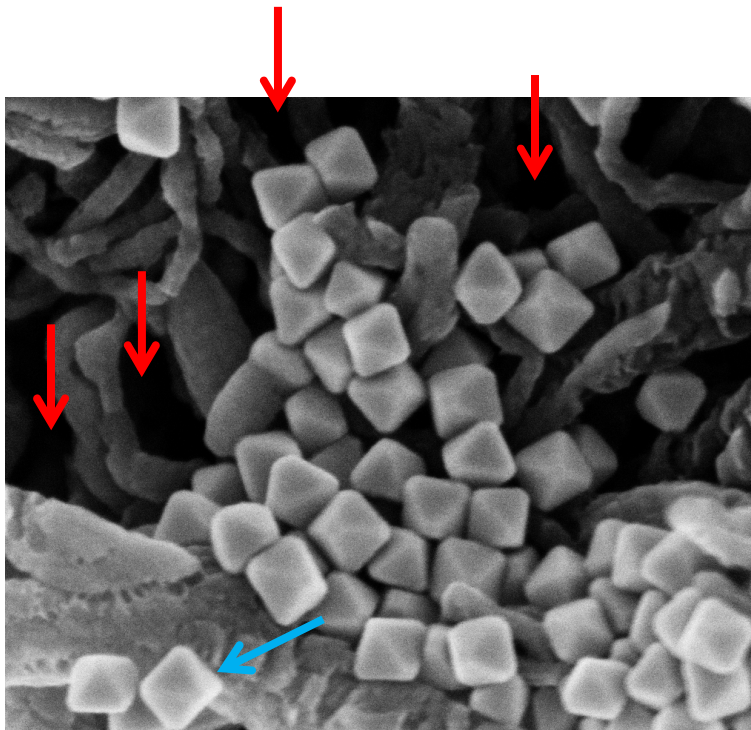


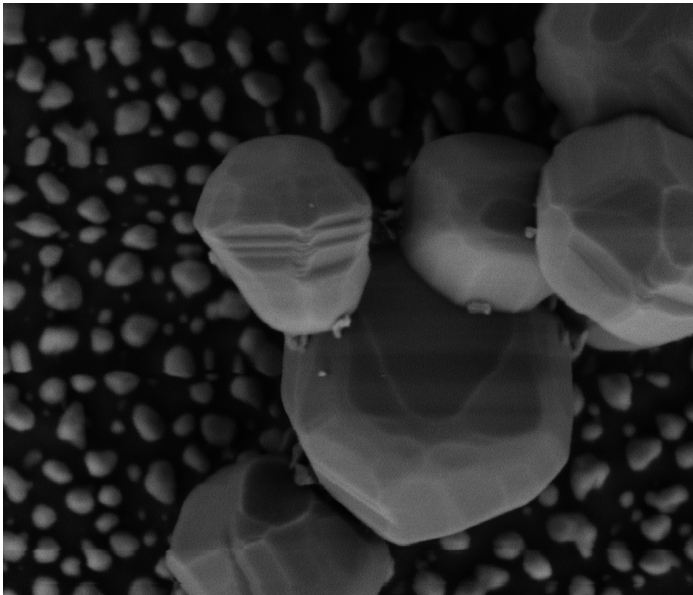
<http://fiji.sc/wiki/index.php/Fiji>

What is a good image..?

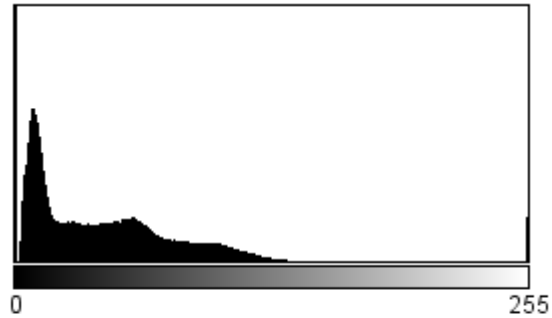


Histogram of 8-bit Tif image



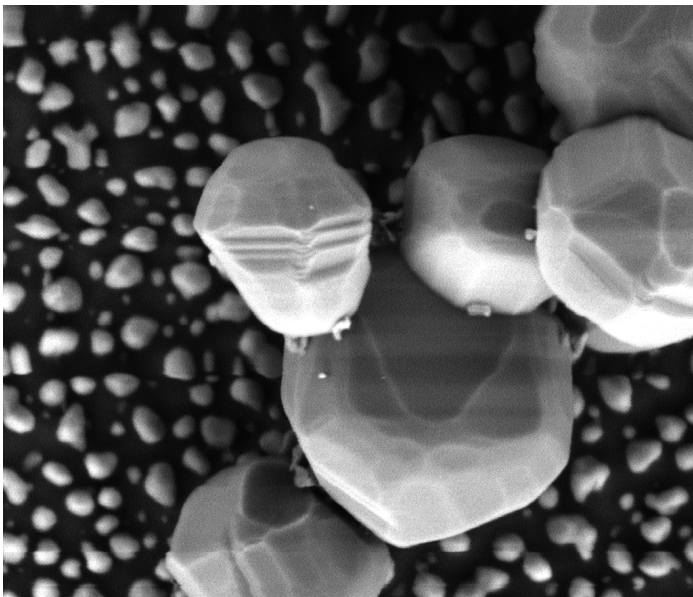


10/30/2008 HV 1.00 kV mag 125 000 x WD 5.0 mm det vCD HFW 1.02 µm tilt -0° 300 nm Magellan 400L

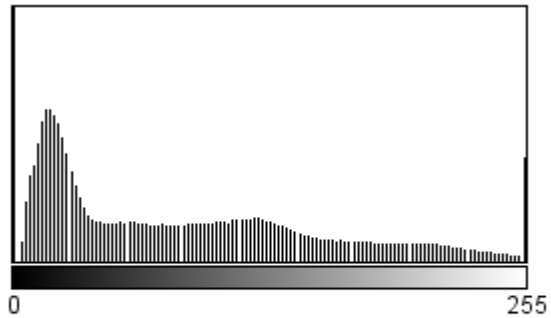


Count: 961536 Min: 0
 Mean: 41.789 Max: 255
 StdDev: 39.195 Mode: 0 (49648)

A little too dark



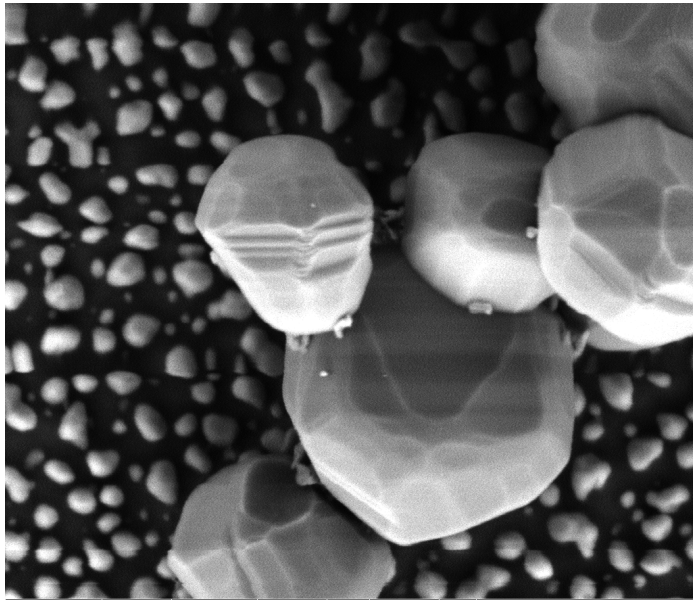
10/30/2008 HV 1.00 kV mag 125 000 x WD 5.0 mm det vCD HFW 1.02 µm tilt -0° 300 nm Magellan 400L



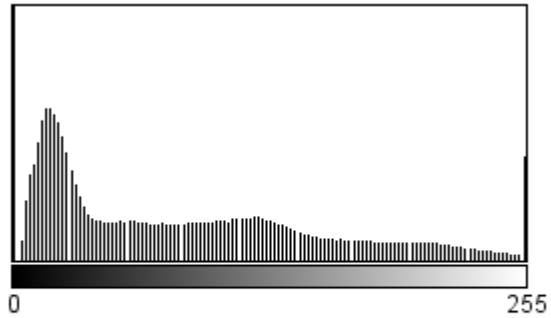
Count: 961536 Min: 0
 Mean: 83.493 Max: 255
 StdDev: 70.754 Mode: 0 (49648)

Adjusted
 contrast/brightness
 (after recording)

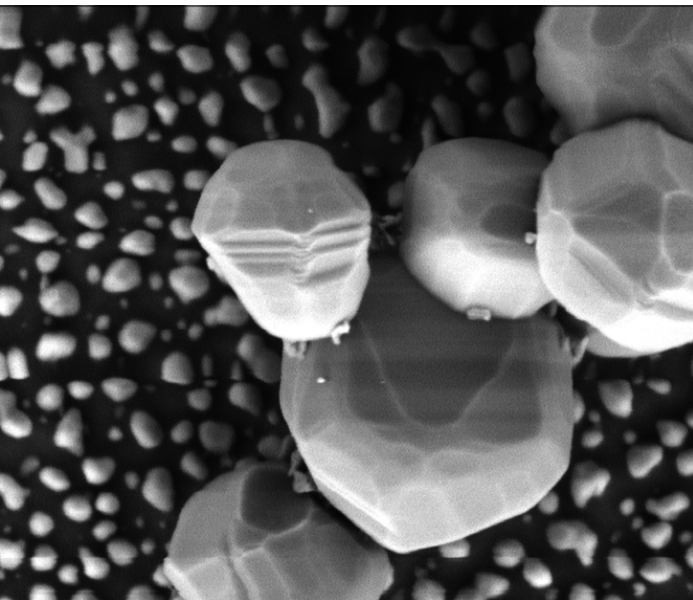
missing grey levels !



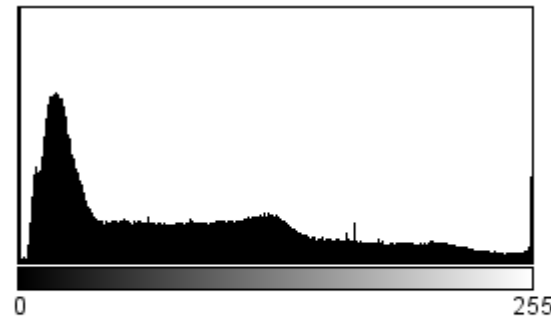
10/30/2008 HV 1.00 kV mag 125 000 x WD 5.0 mm det vCD HFW 1.02 μm tilt -0° 300 nm Magellan 400L



Count: 961536 Min: 0
 Mean: 83.493 Max: 255
 StdDev: 70.754 Mode: 0 (49648)

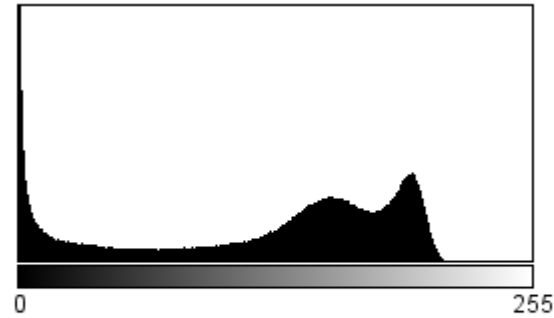
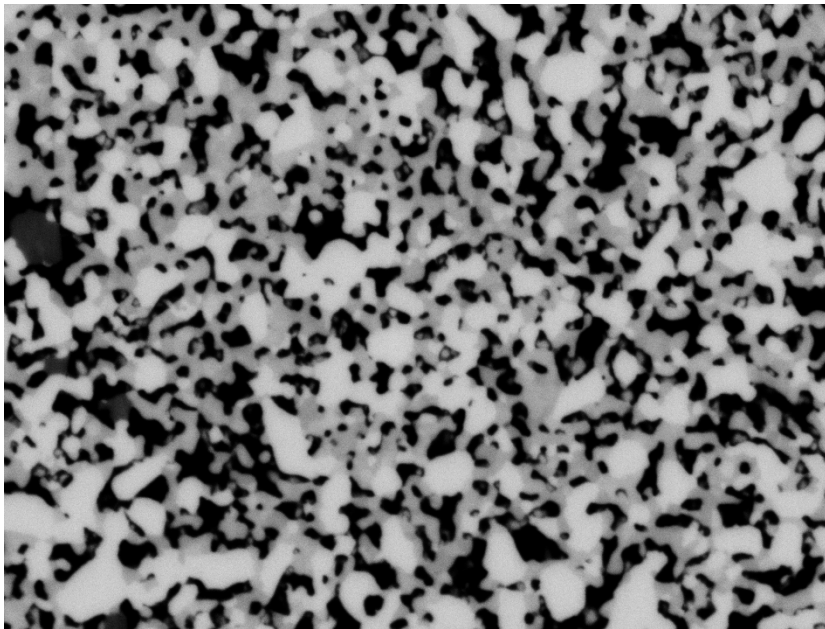


10/30/2008 HV 1.00 kV mag 125 000 x WD 5.0 mm det vCD HFW 1.02 μm tilt -0° 300 nm Magellan 400L

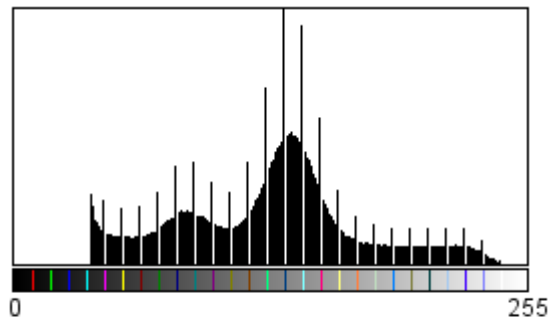


Count: 408258 Min: 0
 Mean: 83.562 Max: 255
 StdDev: 69.314 Mode: 0 (16913)

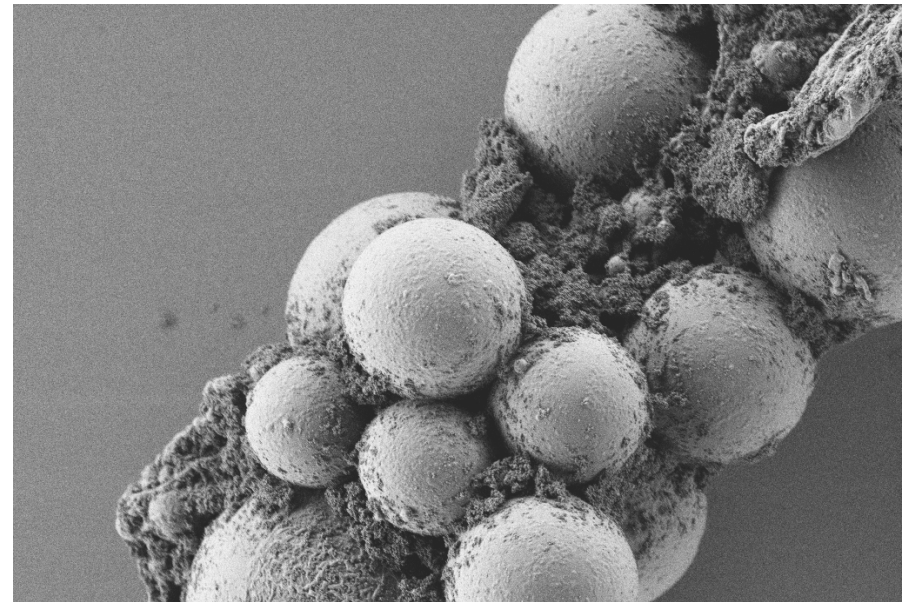
After resampling the image (inventing pixels)



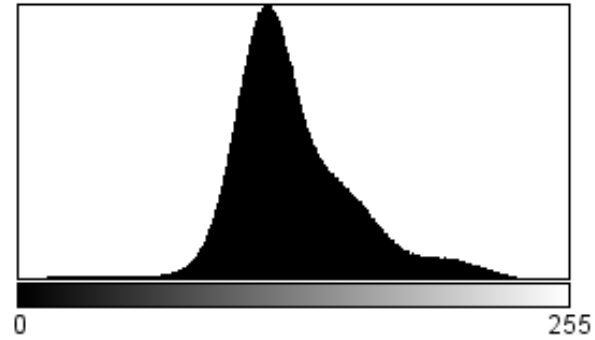
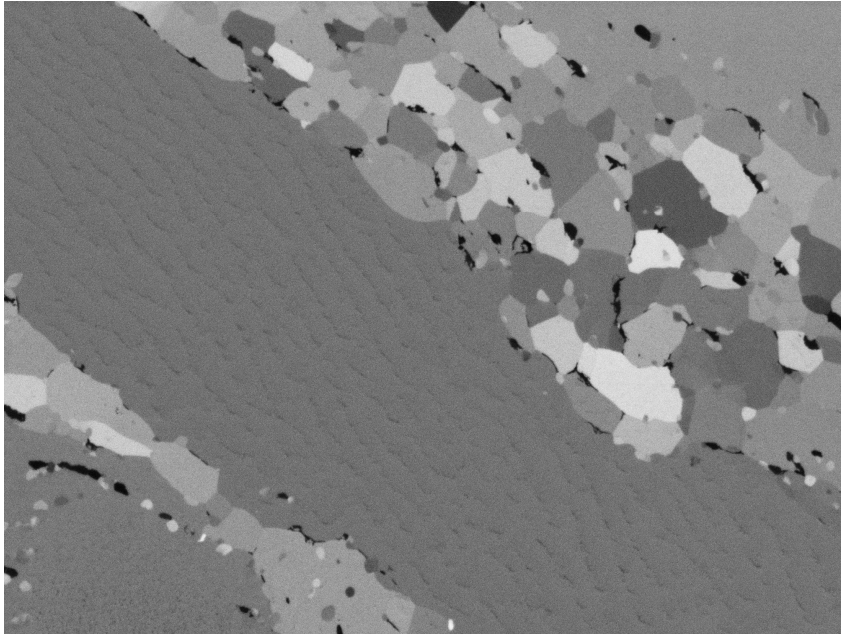
Count: 786432 Min: 0
 Mean: 116.526 Max: 220
 StdDev: 70.839 Mode: 0 (61194)



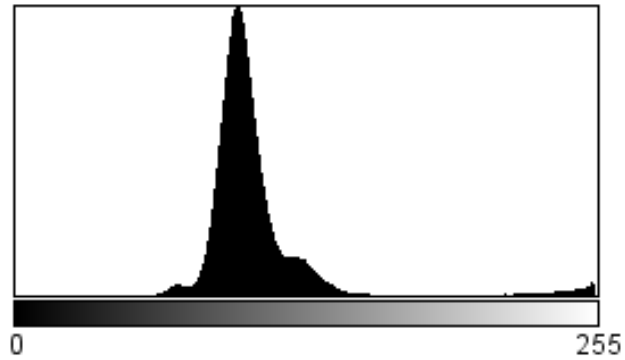
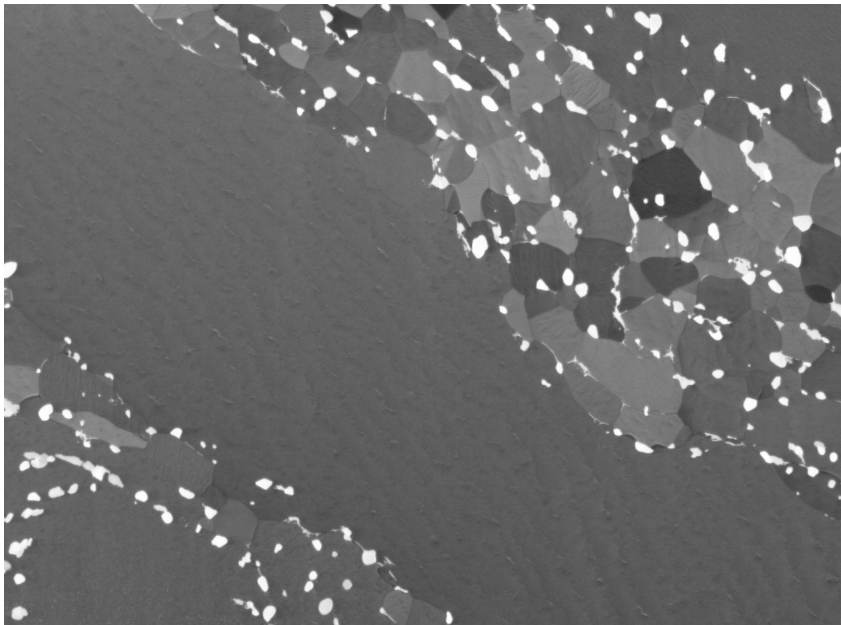
Count: 674674 Min: 37
 Mean: 127.019 Max: 252
 StdDev: 45.457 Mode: 134 (19589)



10 μ m EHT = 0.50 kV Signal A = SEI Width = 76.94 μ m Date :28 Oct 2008
 WD = 2.1 mm Tilt Angle = 36.0 $^{\circ}$ Pixel Size = 75.1 nm Time :13:59:11
 Mag = 1.53 K X Aperture Size = 30.00 μ m File Name = test_02.tif

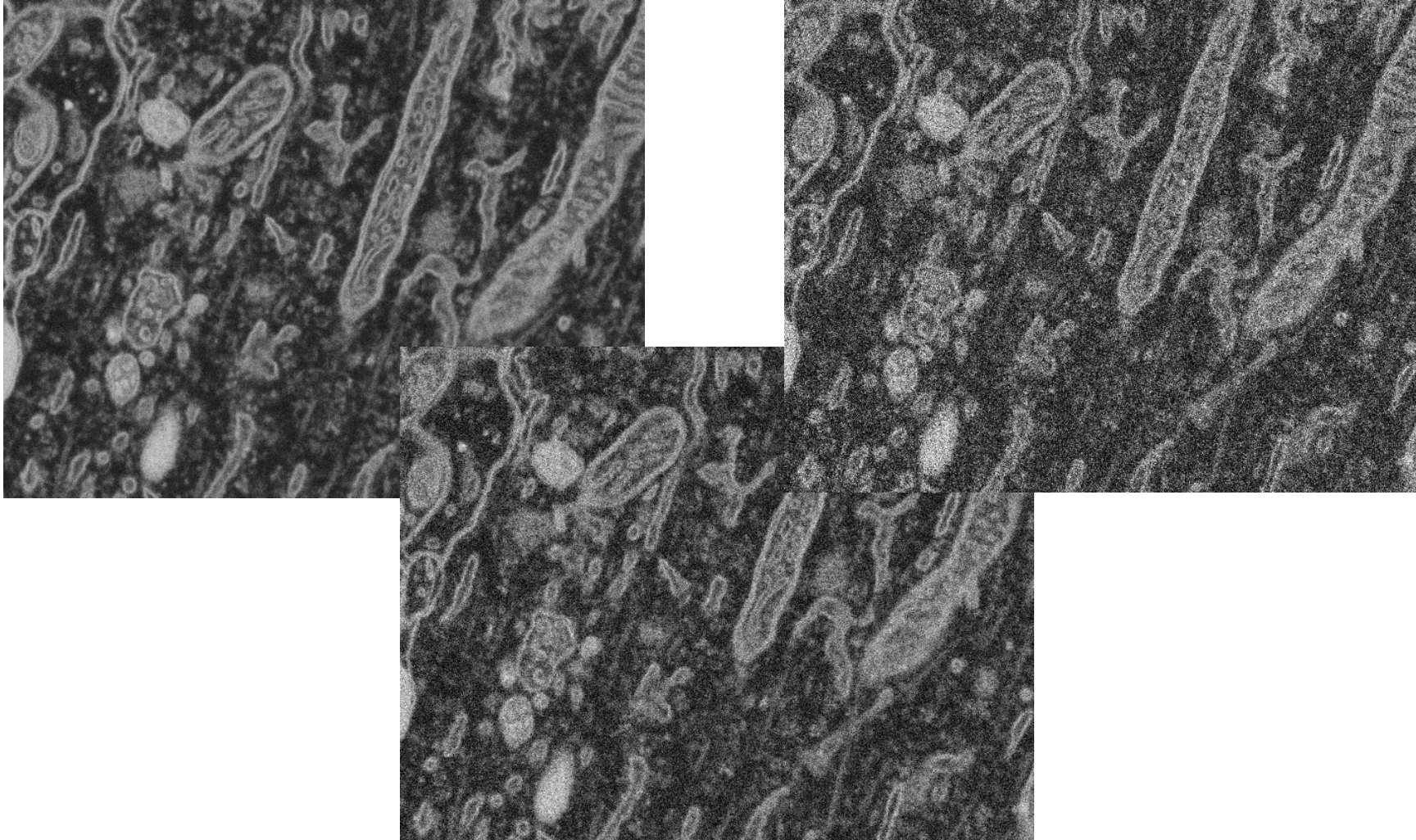


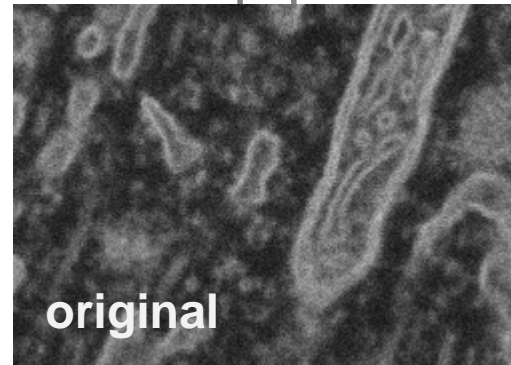
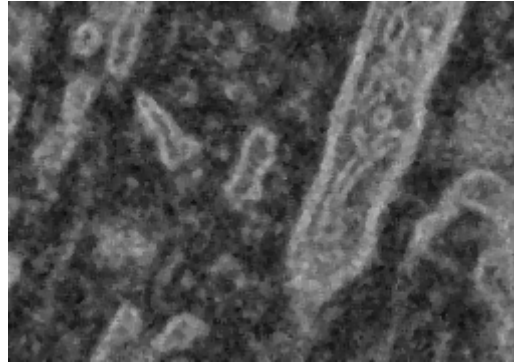
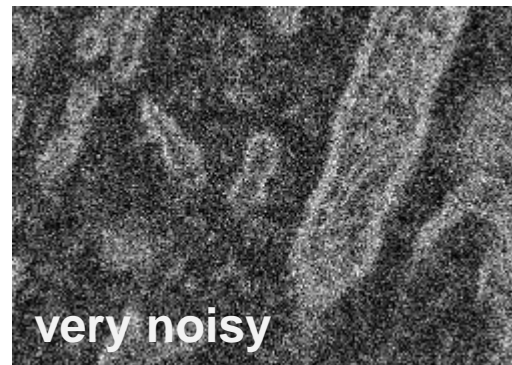
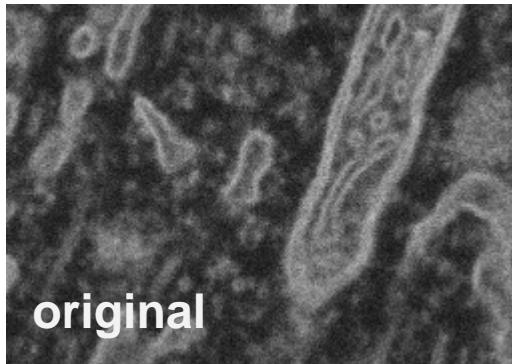
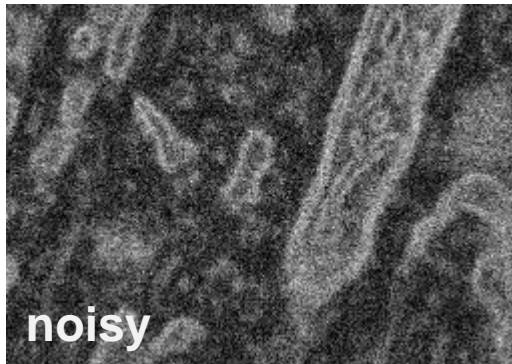
| | |
|----------------|-------------------|
| Count: 2946168 | Min: 1 |
| Mean: 127.193 | Max: 251 |
| StdDev: 29.568 | Mode: 115 (57613) |



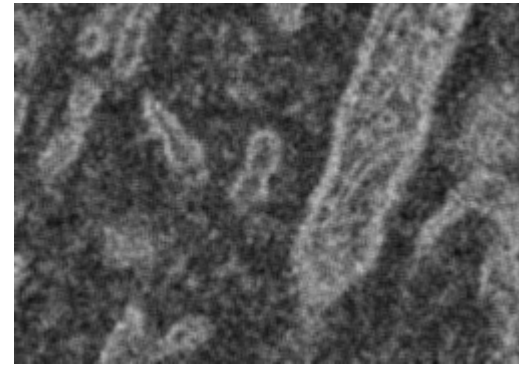
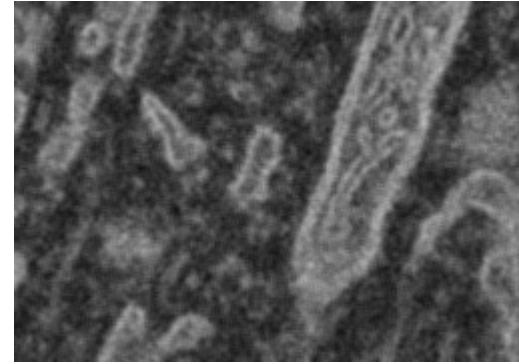
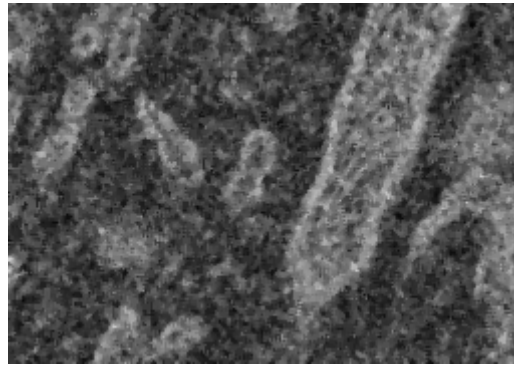
| | |
|----------------|-------------------|
| Count: 3145728 | Min: 14 |
| Mean: 106.215 | Max: 254 |
| StdDev: 28.778 | Mode: 97 (133319) |

dealing with noise





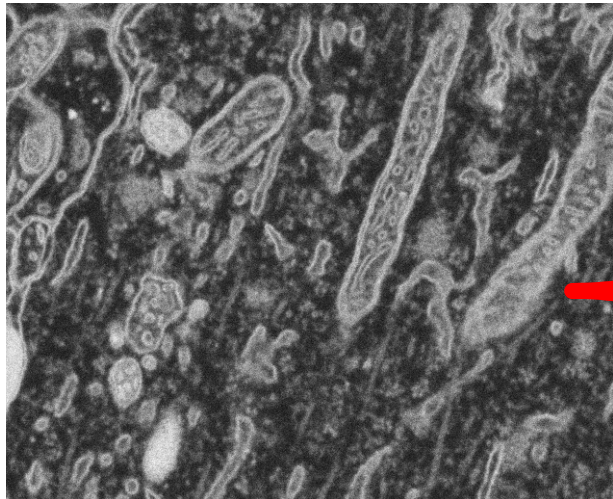
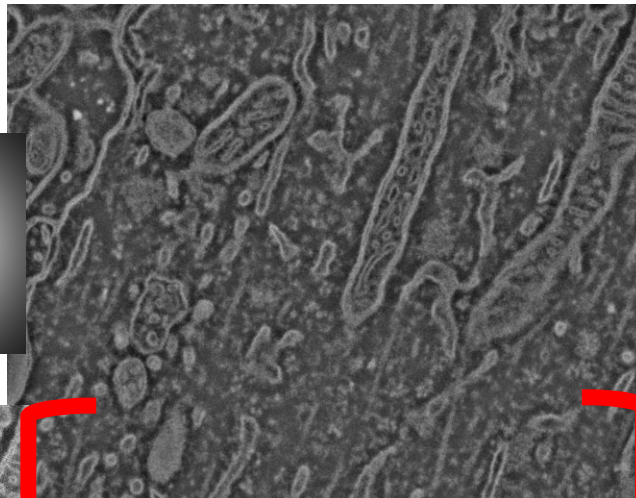
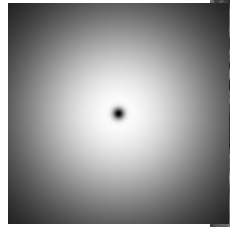
smooth



Filters....

- Median
- Gaussian blur ...reduce in general the resolution
- Mean They take into account the neighbourhood of a pixel
- FFT (fast fourier transform) filter (bandpass)

FFT bandpass filter



High-pass, Small size details
(high space frequency)

Low-pass, Large size details
(low space frequency)

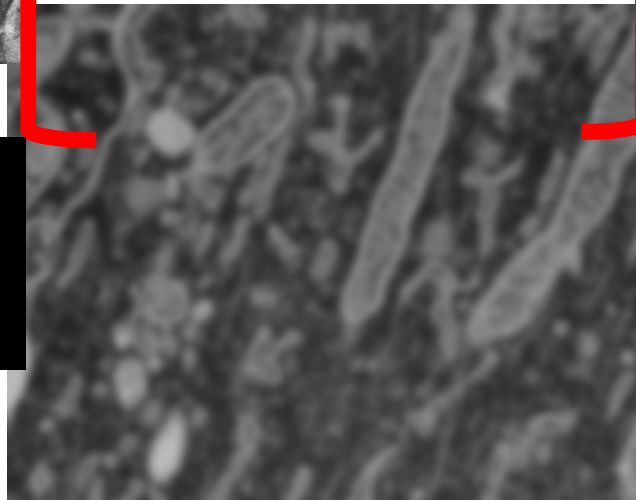
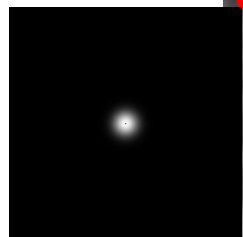
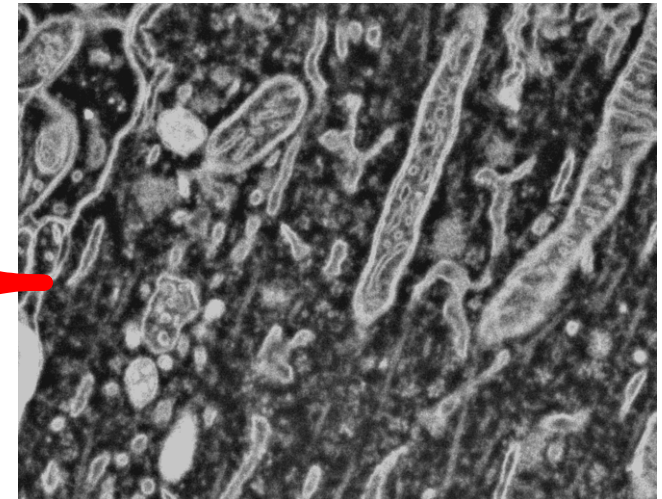
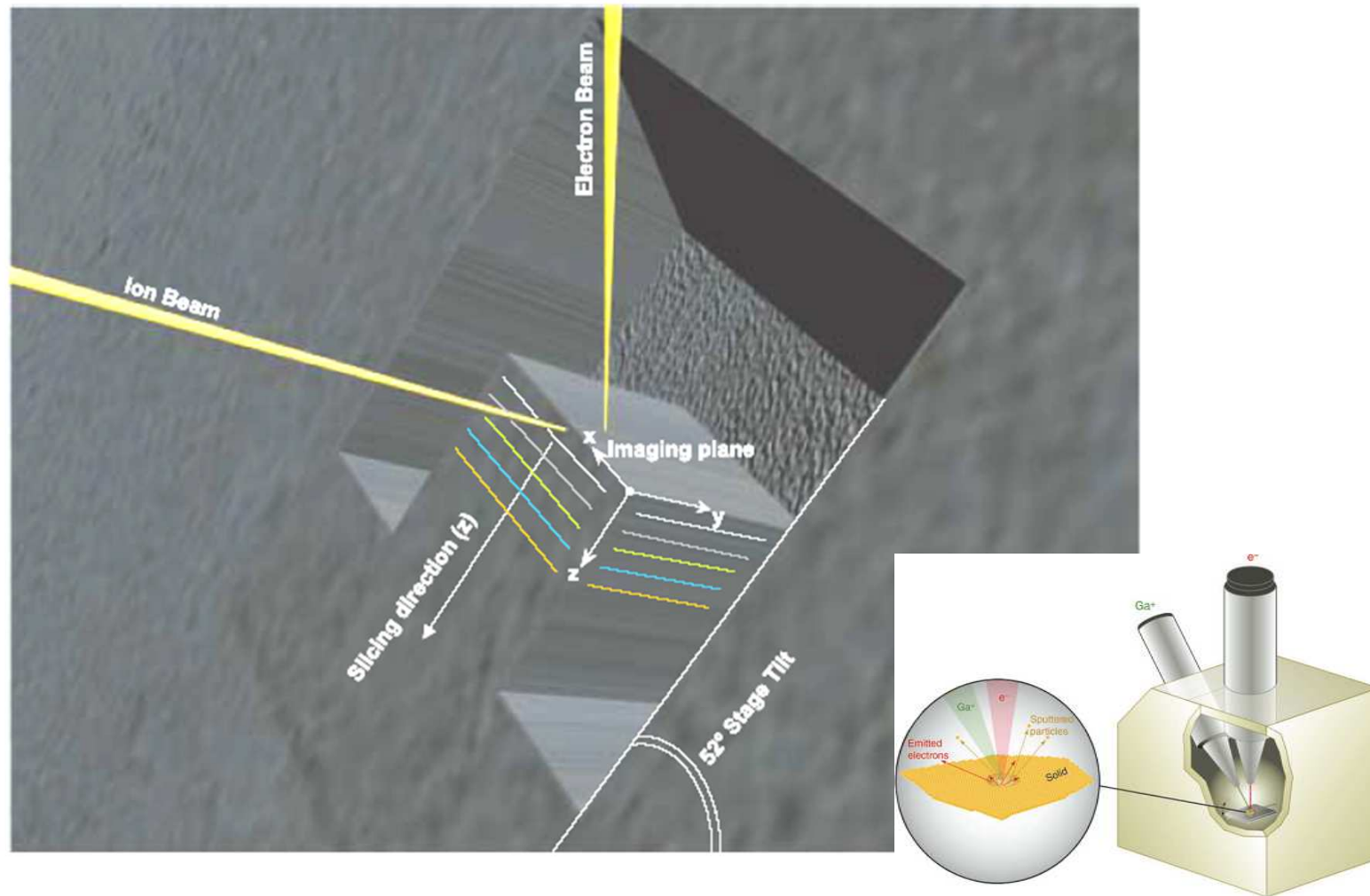


Image processing in 2D

- Best: start with a “good” image:
 - equalized histogram, use all 256 grey levels
 - Reduce noise during acquisition
- Carefully use filters
 - Filtering means “mixing-in” of information from neighbouring pixels: loss of resolution
 - try several to see which one works best
 - do resampling only in the very end

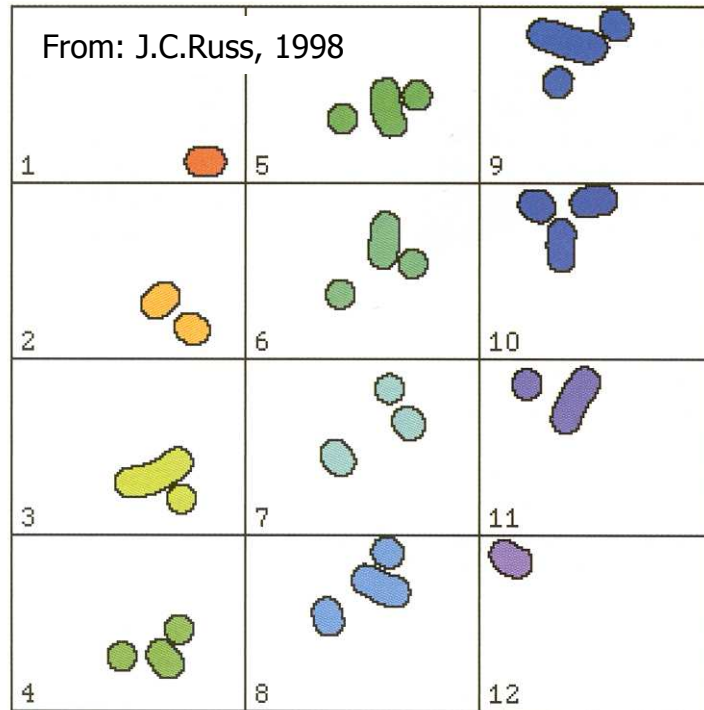
3D imaging with a FIB/SEM

FIB Nanotomography



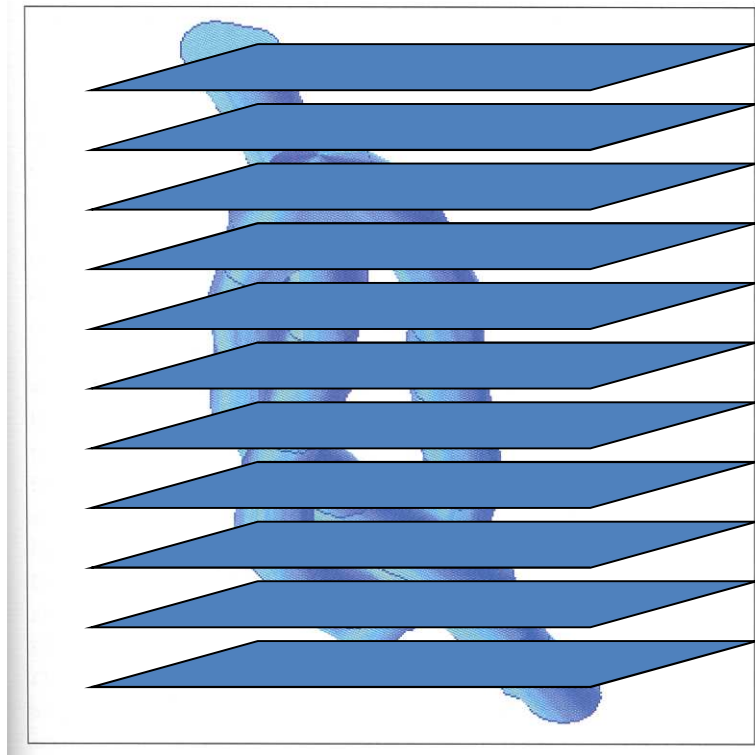
3D Microscopy

Problem of serial sectioning:
3D-reconstruction of disordered microstructures



2D Volume fraction

3D
→

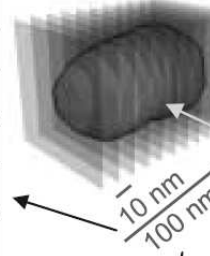


?? Nr of particles ??
?? Shape ??

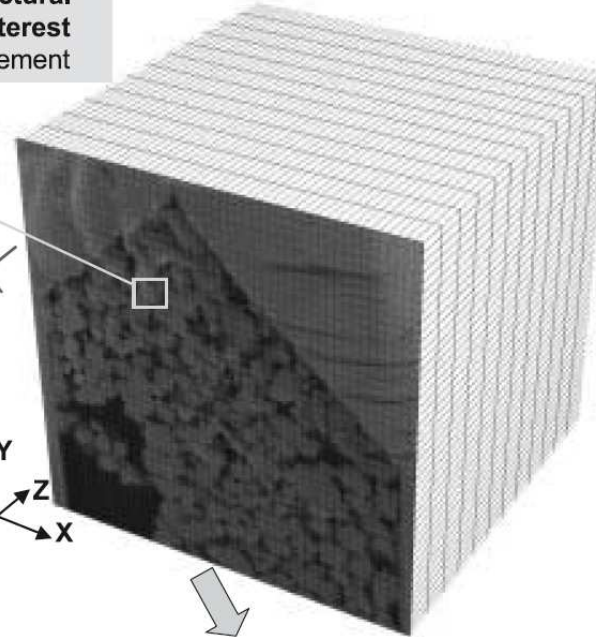
Voxel, Resolution, Pixel size

| Ion-Beam | | | |
|------------------|-----|-----------------|--------------|
| Smallest element | Mag | Ideal z-spacing | Beam current |
| nm | kX | nm | pA |
| 1000 | 5 | 100 | 500 |
| 500 | 10 | 50 | 300 |
| 300 | 15 | 30 | 100 |
| 200 | 30 | 20 | 50 |
| 100 | 50 | 10 | 50 |

Smallest structural element of interest
10 sections/element



Raw data volume

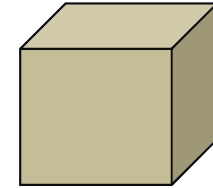


| Electron-Beam | | | | | |
|------------------|-----|--------------|-------------|------------|------|
| Smallest element | Mag | Image width | | Pixel size | |
| | | 1024 pxls, X | 884 pxls, Y | X | Y |
| nm | kX | μm | μm | nm | nm |
| 1000 | 5 | 60.0 | 65.73 | 58.6 | 67.9 |
| 500 | 10 | 30.0 | 32.86 | 29.3 | 33.9 |
| 300 | 15 | 20.0 | 21.91 | 19.5 | 22.6 |
| 200 | 30 | 10.0 | 10.95 | 9.8 | 11.3 |
| 100 | 50 | 6.0 | 5.18 | 5.9 | 6.8 |

| Present example: | x | y | z |
|------------------|----------|------|------------|
| Cube size [μm] | 6.04 | 6.01 | 1.74 |
| Voxel matrix | 1024 | 884 | 105 |
| | [pixels] | | [sections] |
| Voxel size [nm] | 5.9 | 6.8 | 16.6 |

3D FIB/SEM: volume reconstruction

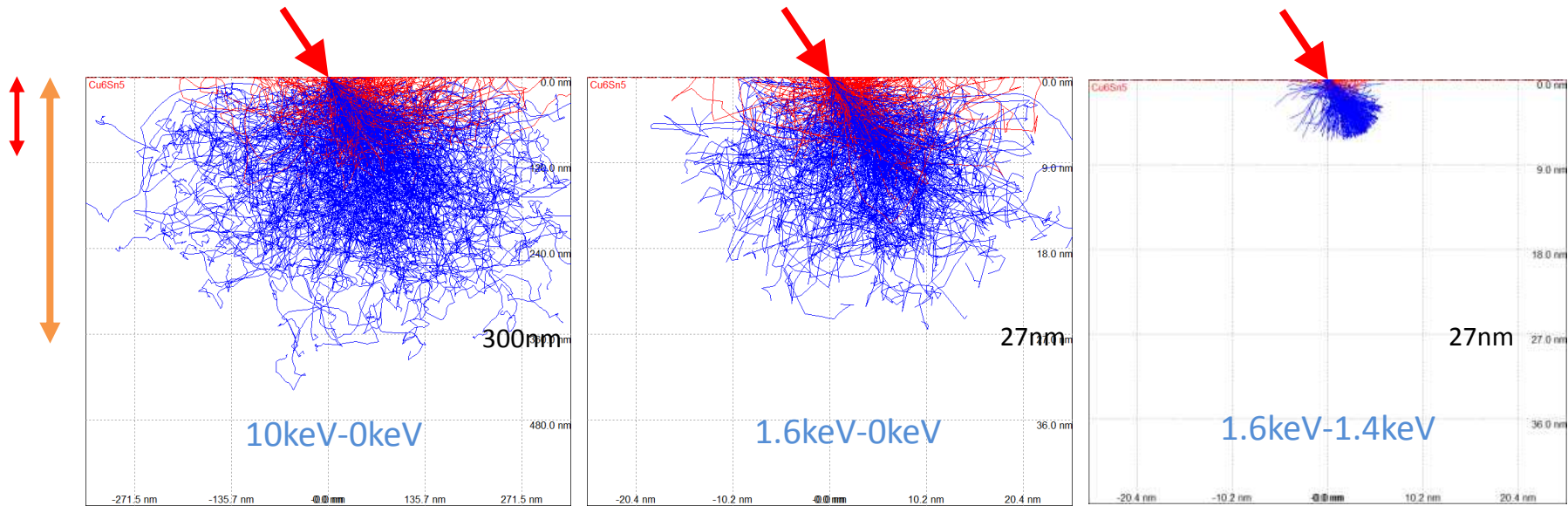
- **Slice thickness (z) = image pixel size (x,y)**
Z dimension \sim X or Y, typical: 10nm, possible 5nm (3nm)
- **Image dimensions / data size (8-bit grey level tiff):**
 - 1024 x 786: 800 slices -> 640 Mb
 - 2048 x 1572: 1600 slices -> 5 Gb
 - 3096 x 2358: 3000 slices -> 21 Gb
- **Acquisition time \sim 1min / slice**
(40-60 slices / hour)
-> high S/N ratio, beam current (1-1.5nA), detector efficiency
- **Dwell times/pixel 5- 15 μ sec.** (detector signal -> 256 grey levels)
- **High throughput:** minimise overhead, no tilting, rotating, drift correction
- **Z- Resolution:** low kV !!!



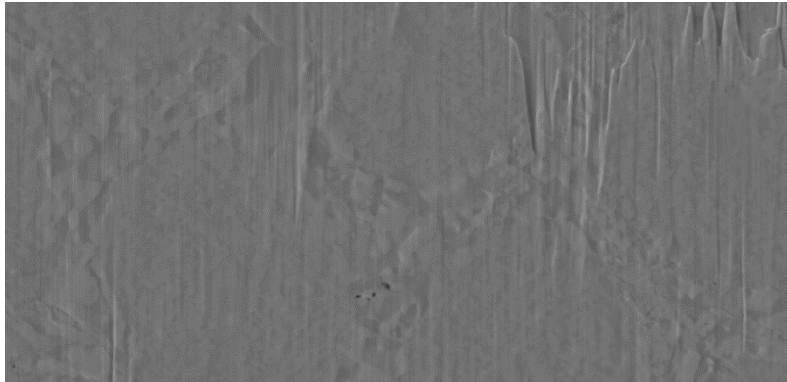
"Leitmotiv"
Isometric voxel size
 $x = y = z$

What is the spatial resolution of BSE electrons ?

Scatter range in Nb₃Sn:

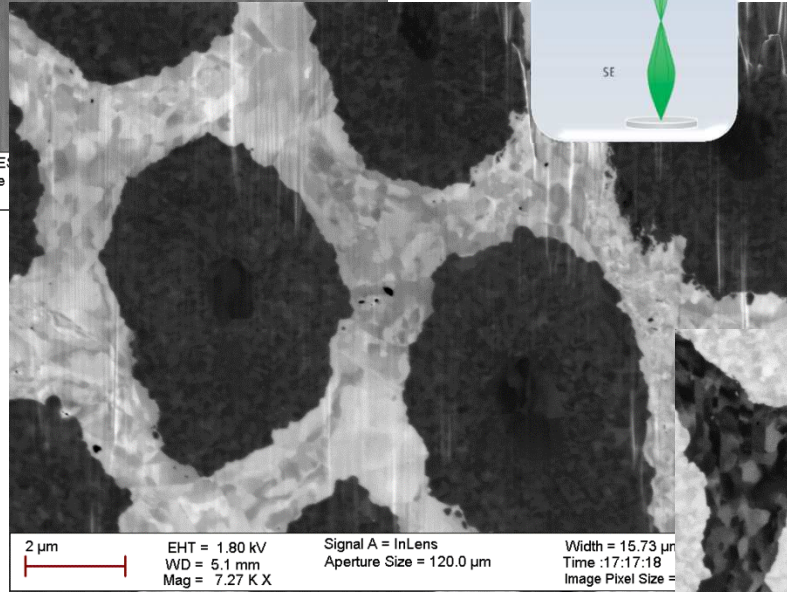
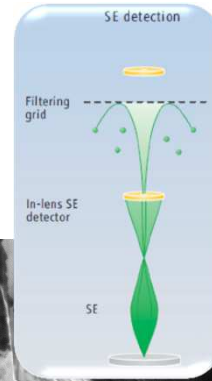


| HT | 10keV | 1.6keV | 1.6keV (low loss, EsB grid at 1.4kV) |
|----------------|-------|--------|---|
| BSE esc. depth | 100nm | 10nm | 2-3nm |
| penetration | 300nm | 20nm | (20nm) Energy selective BS |



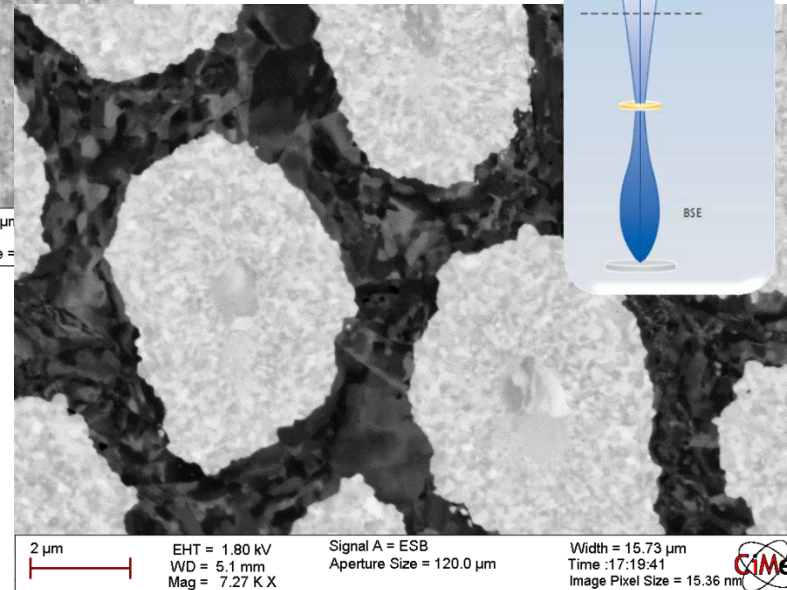
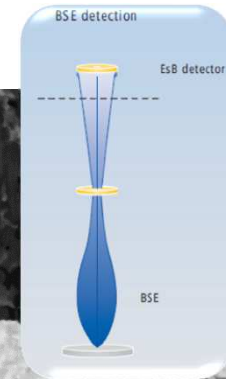
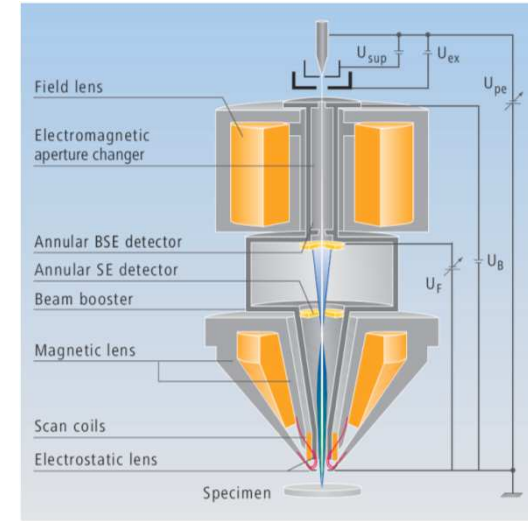
2 μm EHT = 1.80 kV Signal A = SE
 WD = 5.1 mm Aperture Size
 Mag = 7.27 K X

in-chamber ET-detector, SE



2 μm EHT = 1.80 kV Signal A = InLens Width = 15.73 μm
 WD = 5.1 mm Aperture Size = 120.0 μm Time :17:17:18
 Mag = 7.27 K X Image Pixel Size =

in-column "InLens", SE-detector

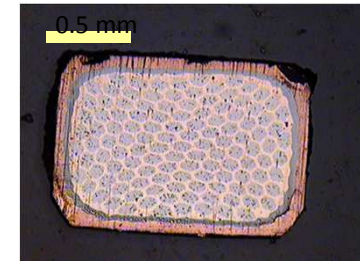
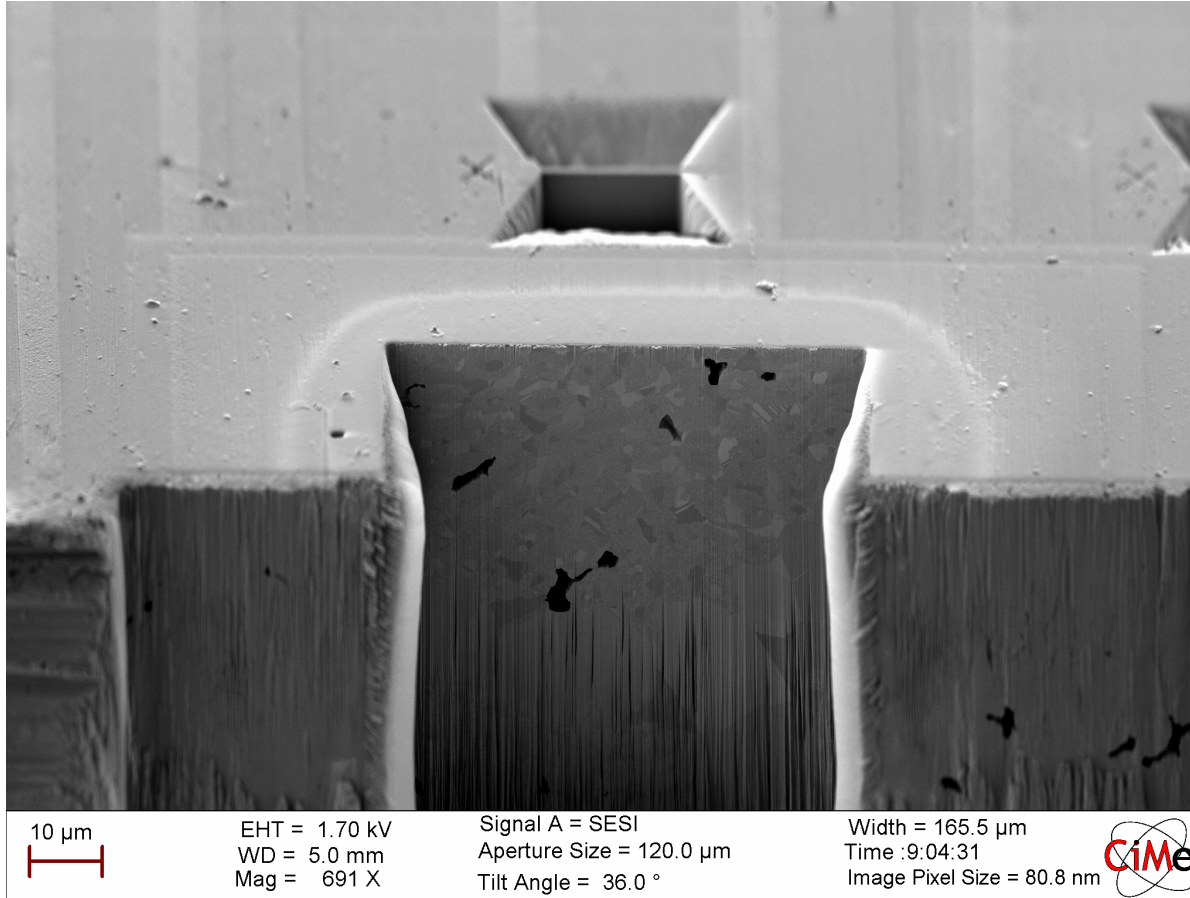


2 μm EHT = 1.80 kV Signal A = EsB Width = 15.73 μm
 WD = 5.1 mm Aperture Size = 120.0 μm Time :17:19:41
 Mag = 7.27 K X Image Pixel Size = 15.36 nm

in-column, "energy-selective" EsB, BSE-detector

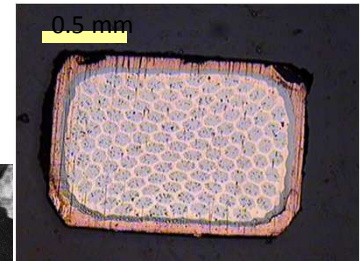
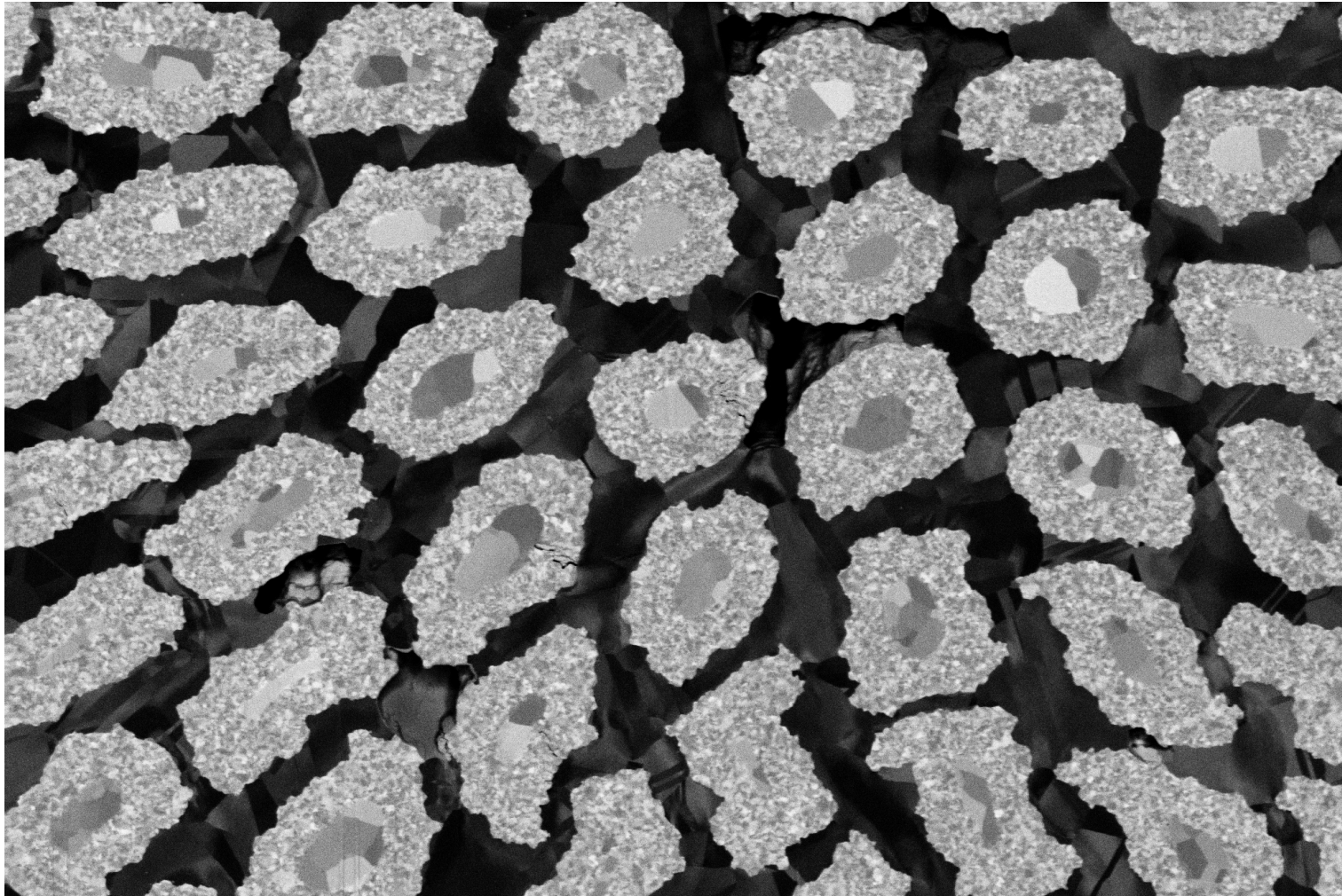
**Low kV imaging:
 acceleration voltage: 1.8 kV
 In-column detectors**

Nb₃Sn multifilament superconducting cable



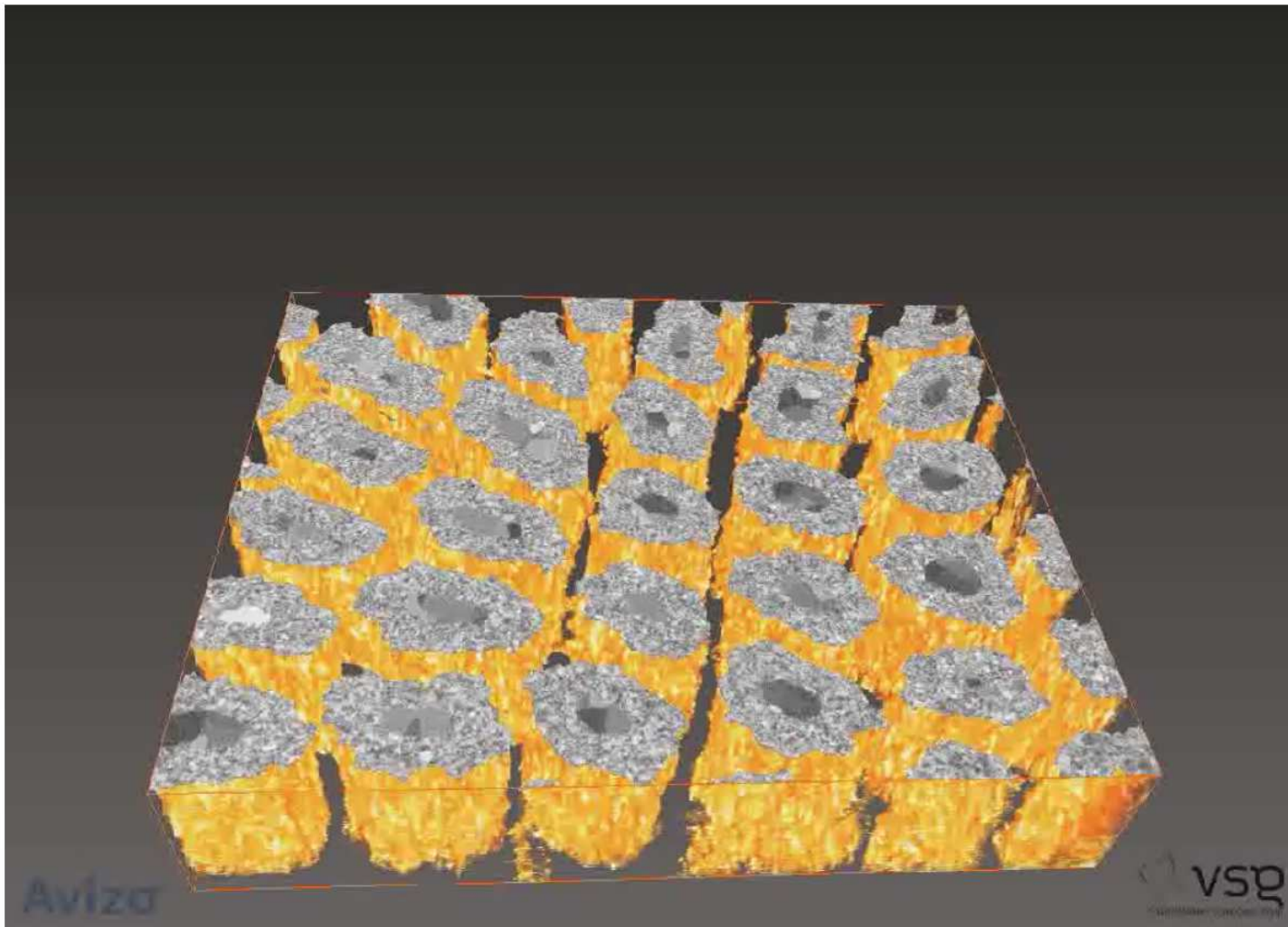
Nb₃Sn superconductor
multifilament cable:
14'000 Nb₃Sn filaments
(diameter ~5μm) in Cu
matrix

Nb₃Sn multifilament superconducting cable



Nb₃Sn superconductor
multifilament cable:
14'000 Nb₃Sn filaments
(diameter ~5μm) in Cu
matrix

1.8kV EsB detector: Materials & orientation contrast



Materials & grain contrast

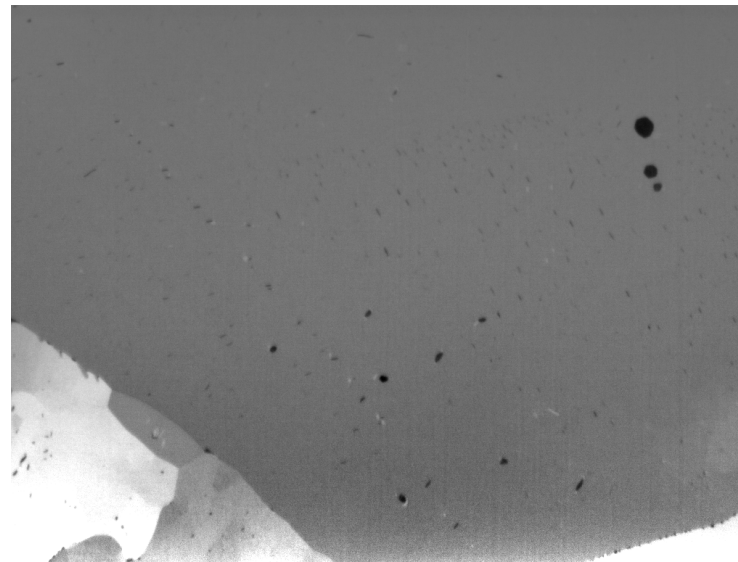
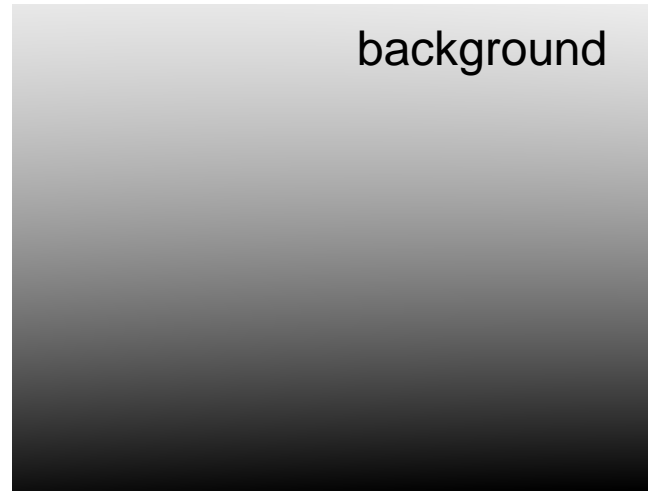
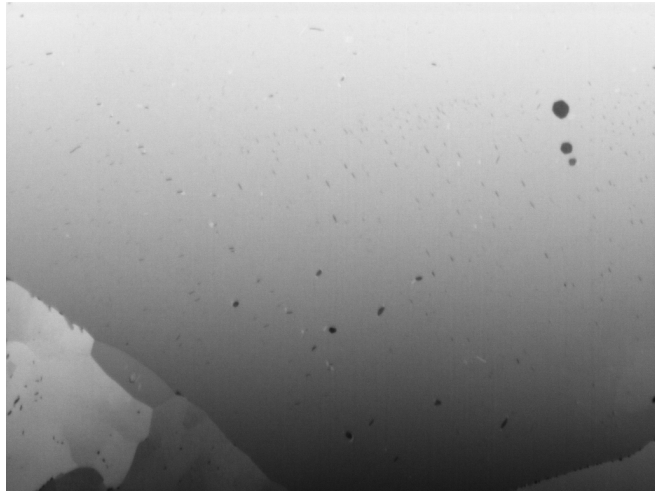
2048x1536x1700, (10x10x10nm voxel),
28hours

Image processing in 3D

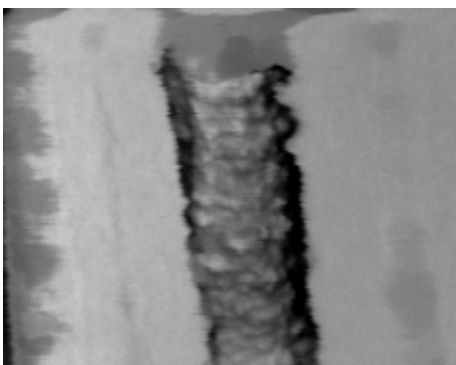
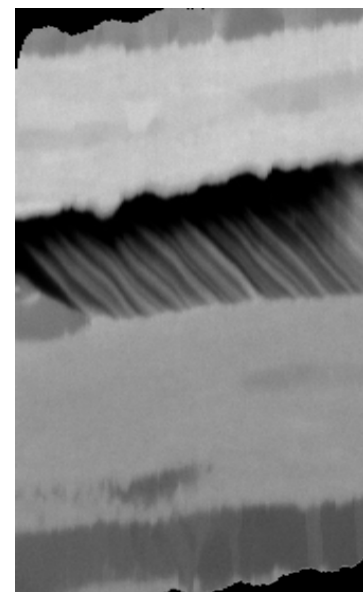
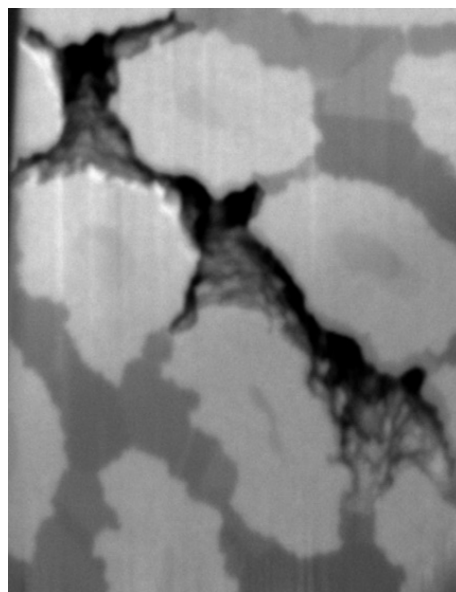
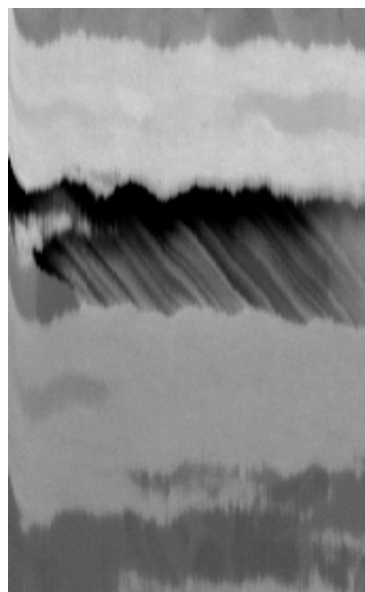
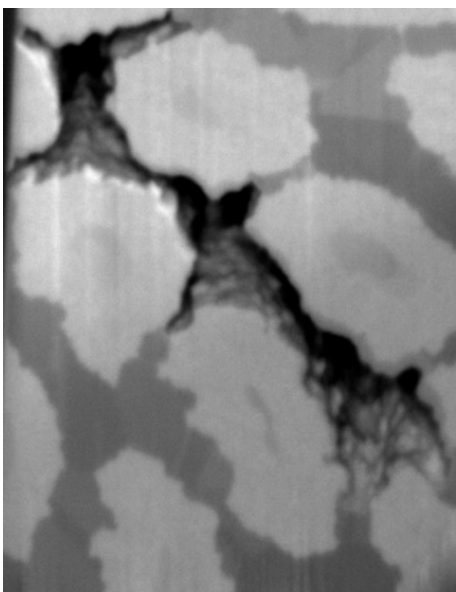
- 2D processing (shading correction)
- Image alignment (registration)
- Cropping (subvolume)
- Noise reduction (2D or 3D)

- Segmentation
- Visualization

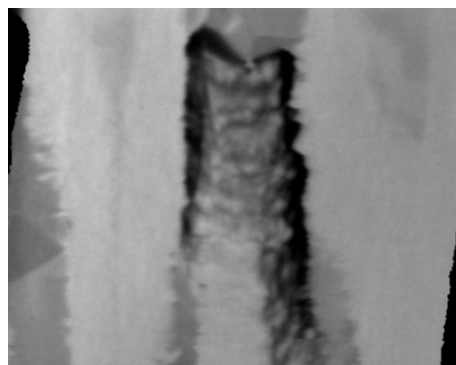
Shading correction



Registration

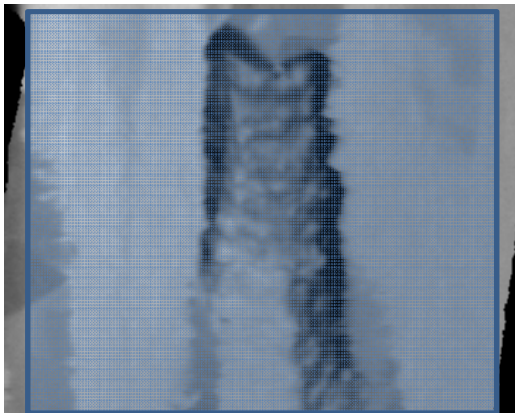
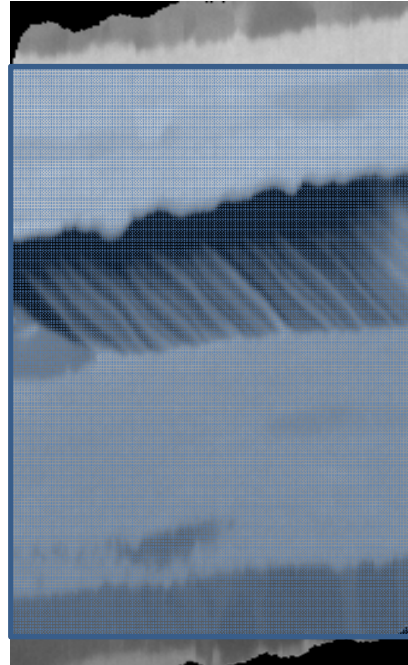
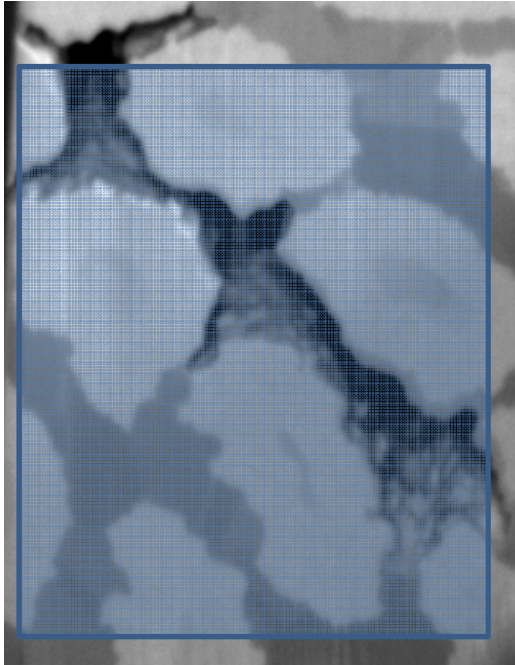


before



after

cropping



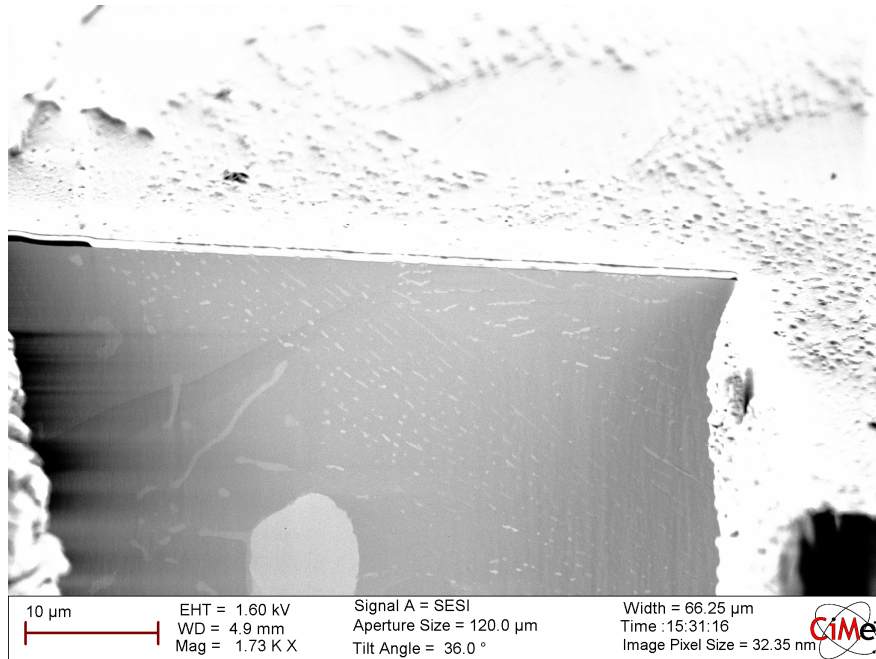
Cutting of the border

Segmentation

- Identifying the different phases/volumes/objects
- Creating binary images/stacks of the different objects

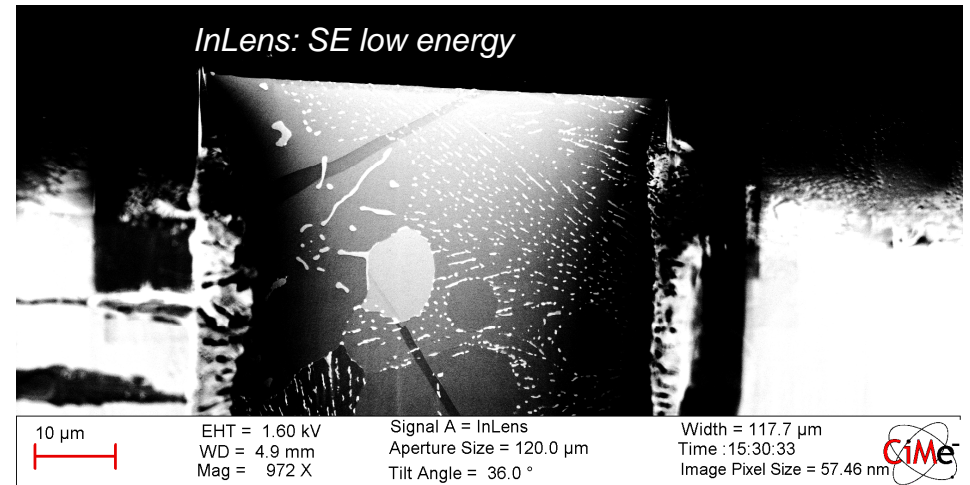
Pb-free solder SnAgCu: "one detector is not enough"

M. Maleki, EPFL-LMAF

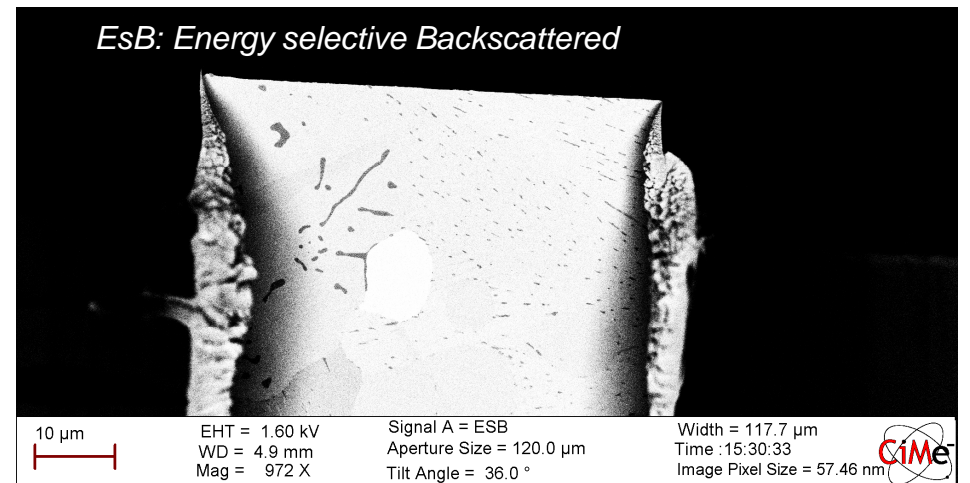


10 µm EHT = 1.60 kV Signal A = SESI Width = 66.25 µm
WD = 4.9 mm Aperture Size = 120.0 µm Time : 15:31:16
Mag = 1.73 K X Tilt Angle = 36.0 ° Image Pixel Size = 32.35 nm

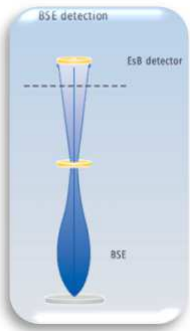
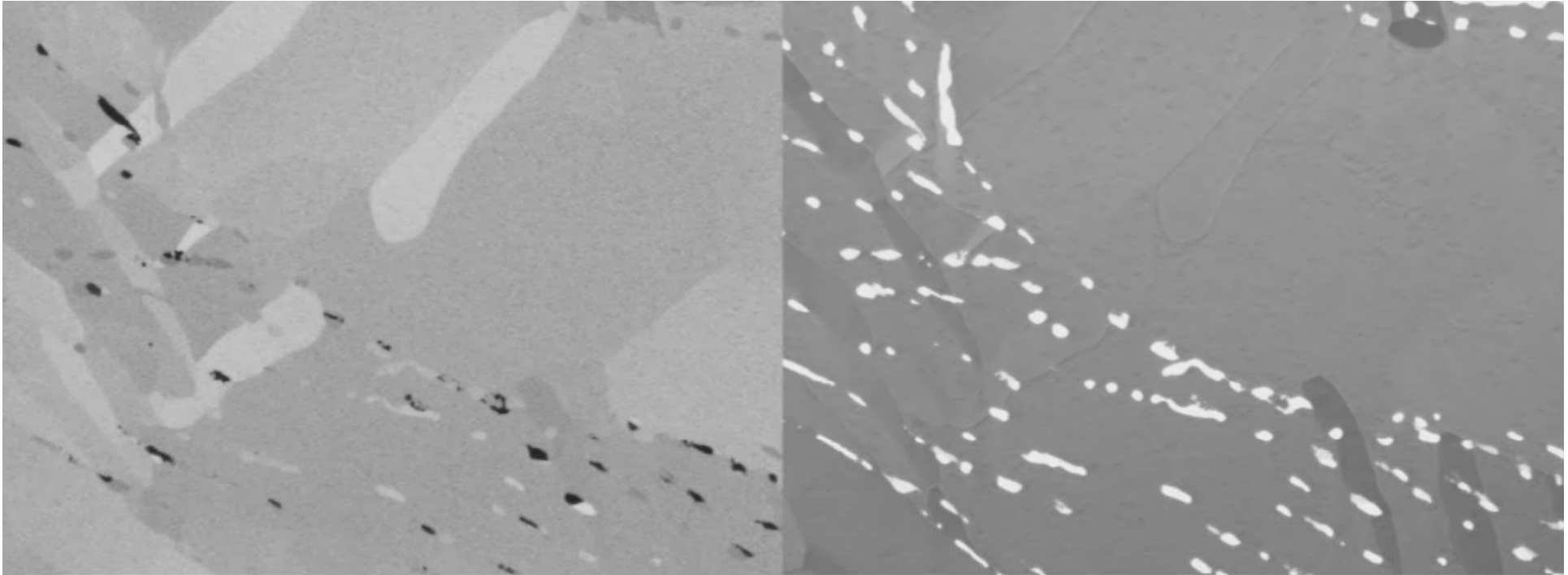
ETD (SE classic)



10 µm EHT = 1.60 kV Signal A = InLens Width = 117.7 µm
WD = 4.9 mm Aperture Size = 120.0 µm Time : 15:30:33
Mag = 972 X Tilt Angle = 36.0 ° Image Pixel Size = 57.46 nm



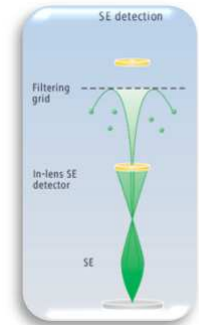
10 µm EHT = 1.60 kV Signal A = ESB Width = 117.7 µm
WD = 4.9 mm Aperture Size = 120.0 µm Time : 15:30:33
Mag = 972 X Tilt Angle = 36.0 ° Image Pixel Size = 57.46 nm

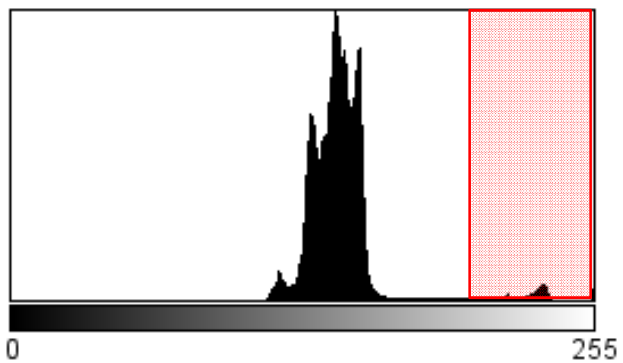
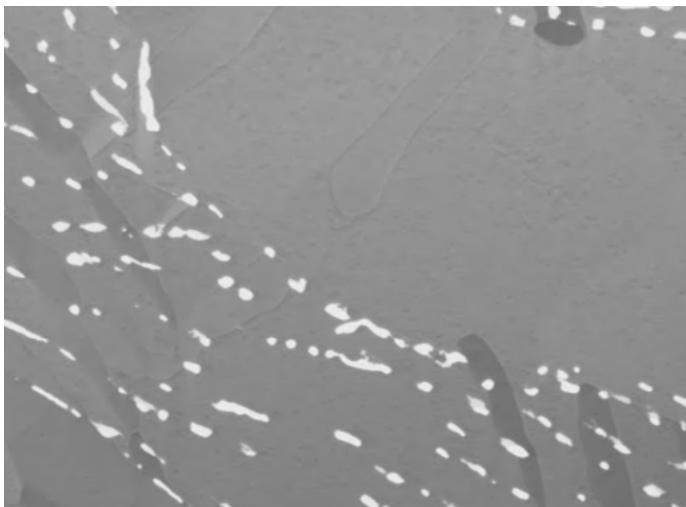


EsB

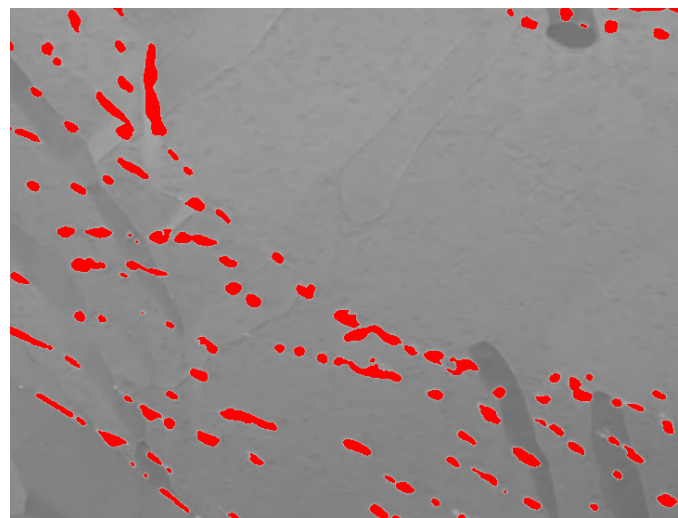
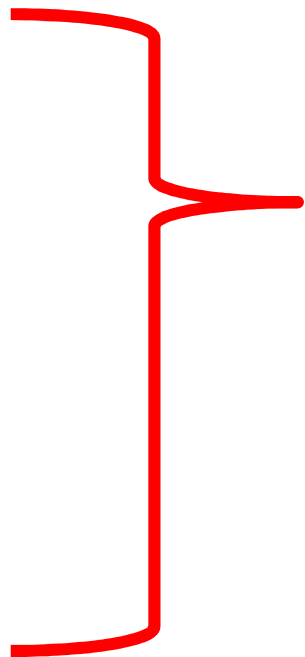
10x10x10nm voxel size, 2048x1536x2000
2 images (2x3Mb) / slice ...! (DUAL Channel !)
1.6keV, EsB & InLens-SE detector
12Gb data

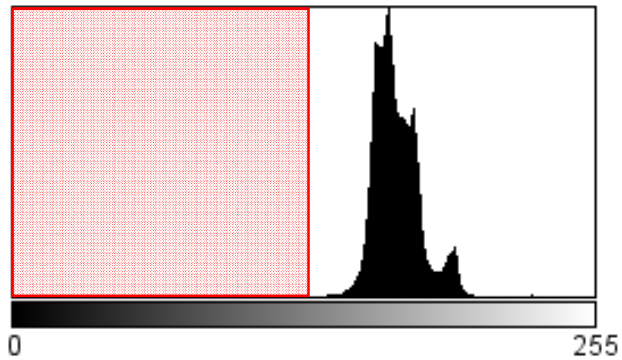
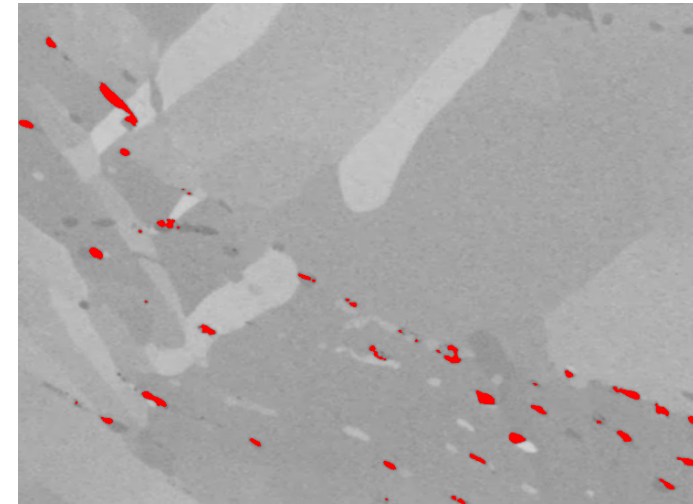
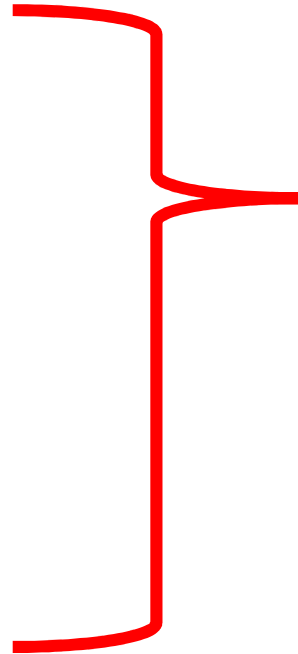
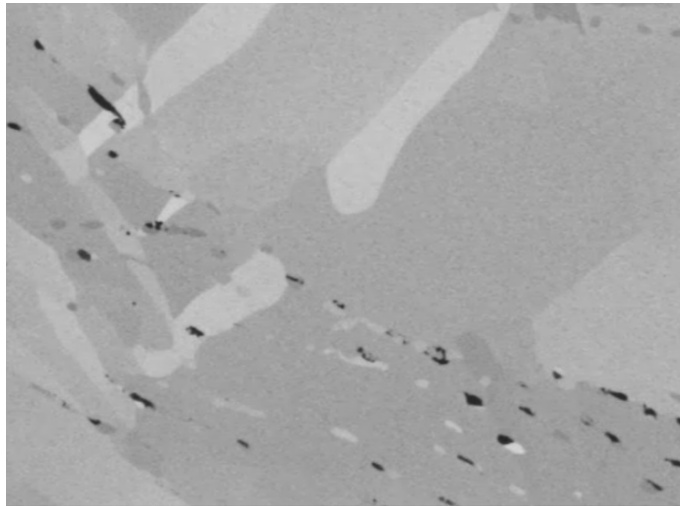
InLens SE



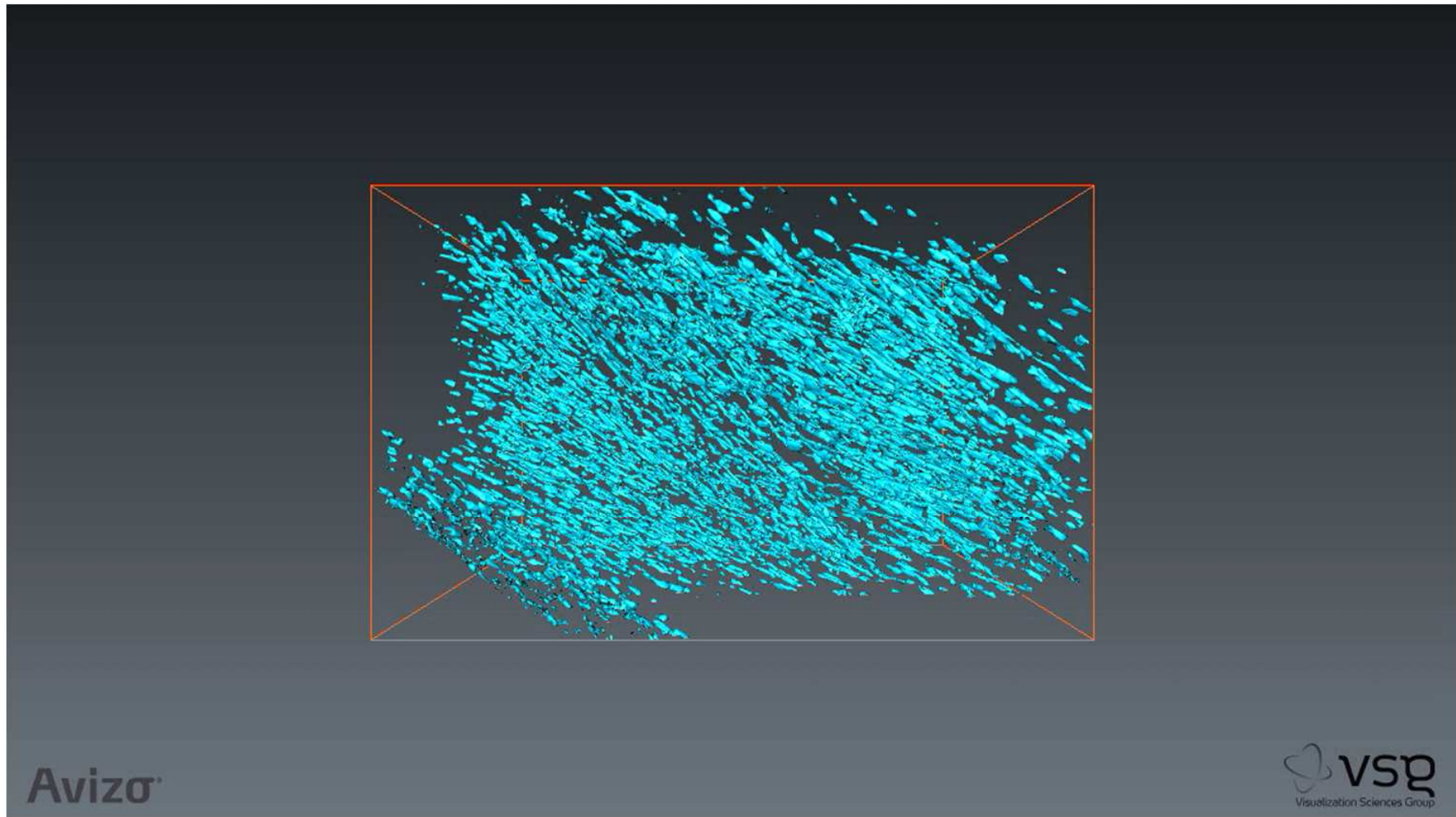


Count: 411792 Min: 101
Mean: 145.445 Max: 255
StdDev: 18.865 Mode: 142 (18748)





Count: 411792 Min: 36
Mean: 167.731 Max: 236
StdDev: 13.290 Mode: 165 (20261)



Phase 1. Dark in EsB image, White in SE-InLens

10x10x10nm voxel size, 2048x1536x2000 pixel/slices

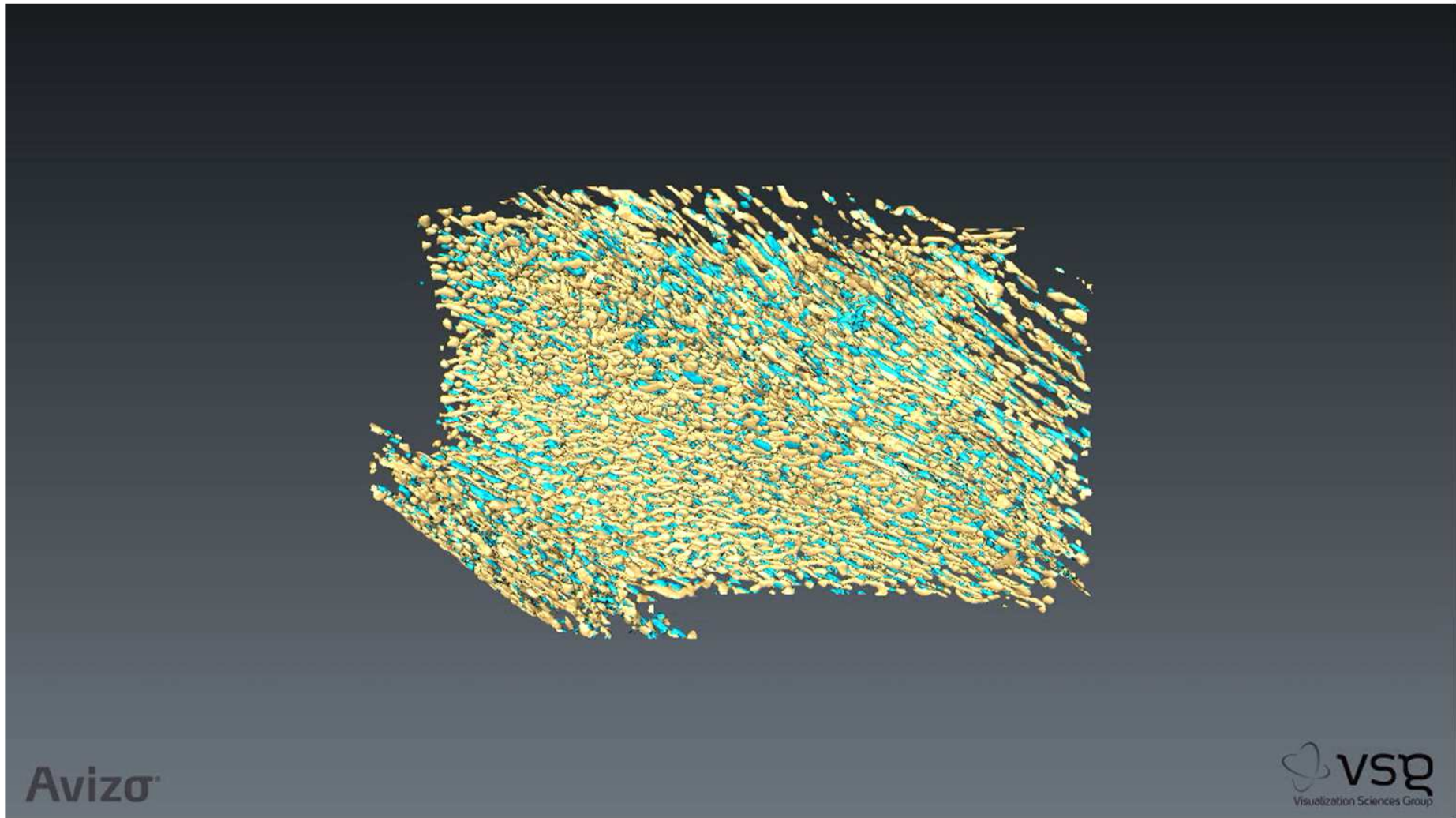
2 images (3Mb) / slice 12Gb data



Phase 2: White in SE-InLens - Dark in EsB image

10x10x10nm voxel size, 2048x1536x2000 pixel/slices

2 images (3Mb) / slice 12Gb data



Avizo[®]

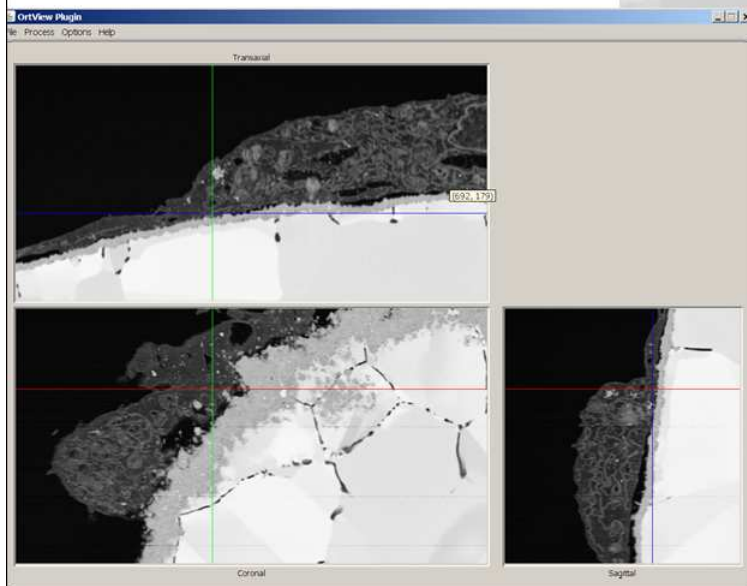
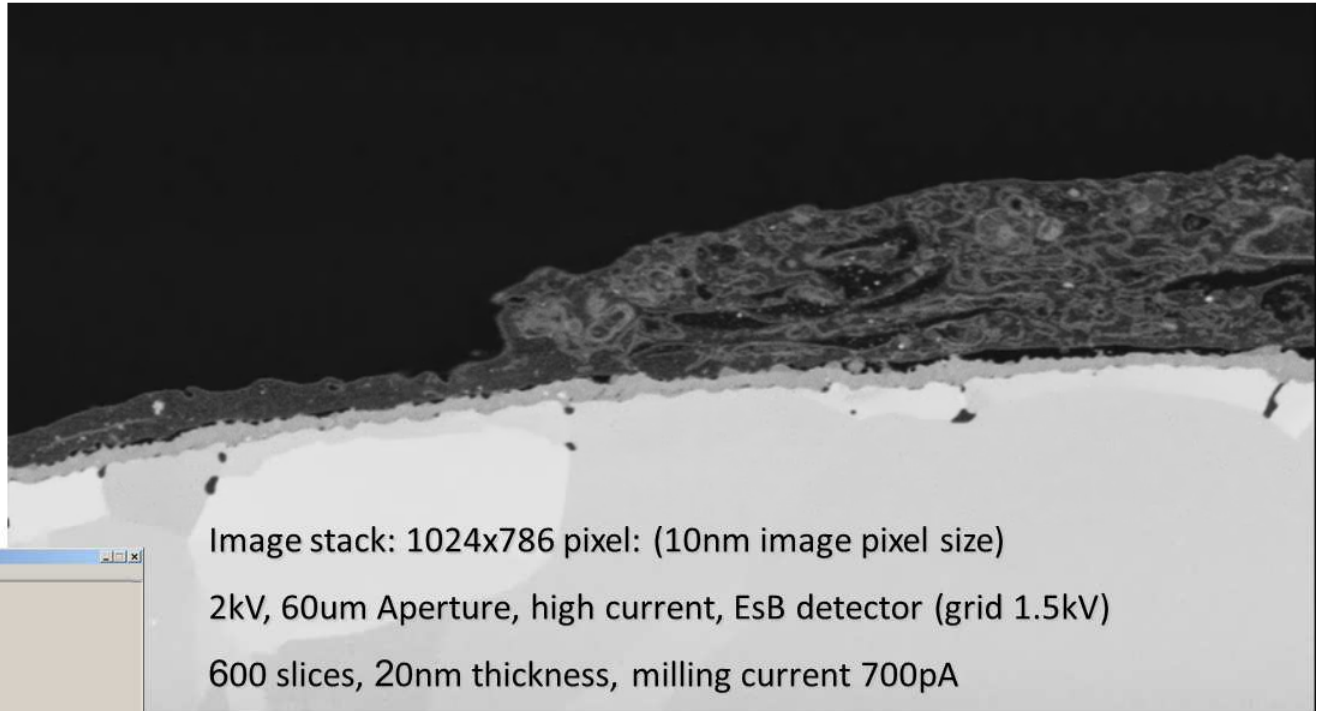
 **vsg**
Visualization Sciences Group

Segmentation tools for life-science

Problem:

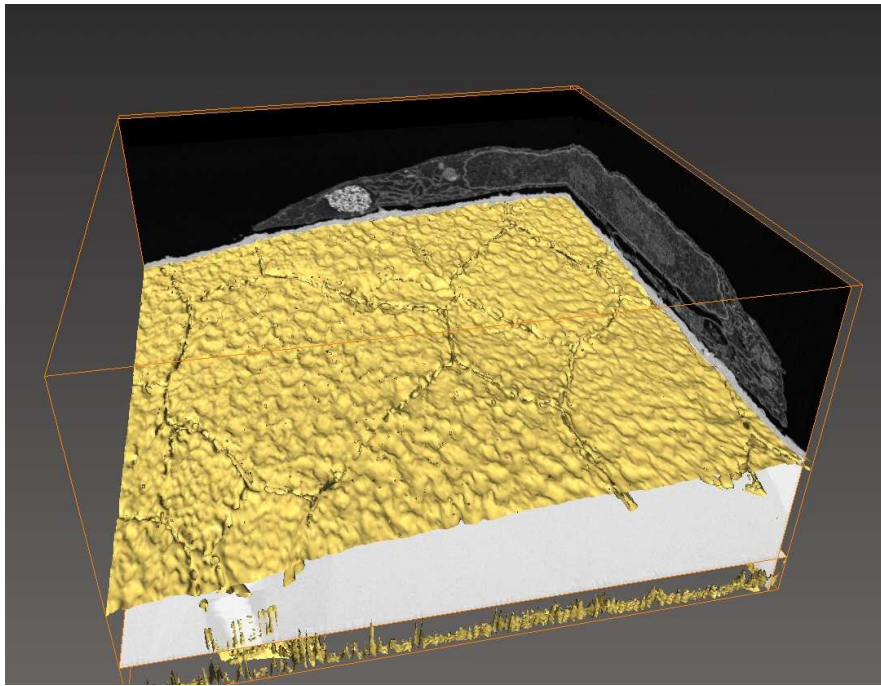
- Structure (shape) determines function of objects

Single cell on substrate

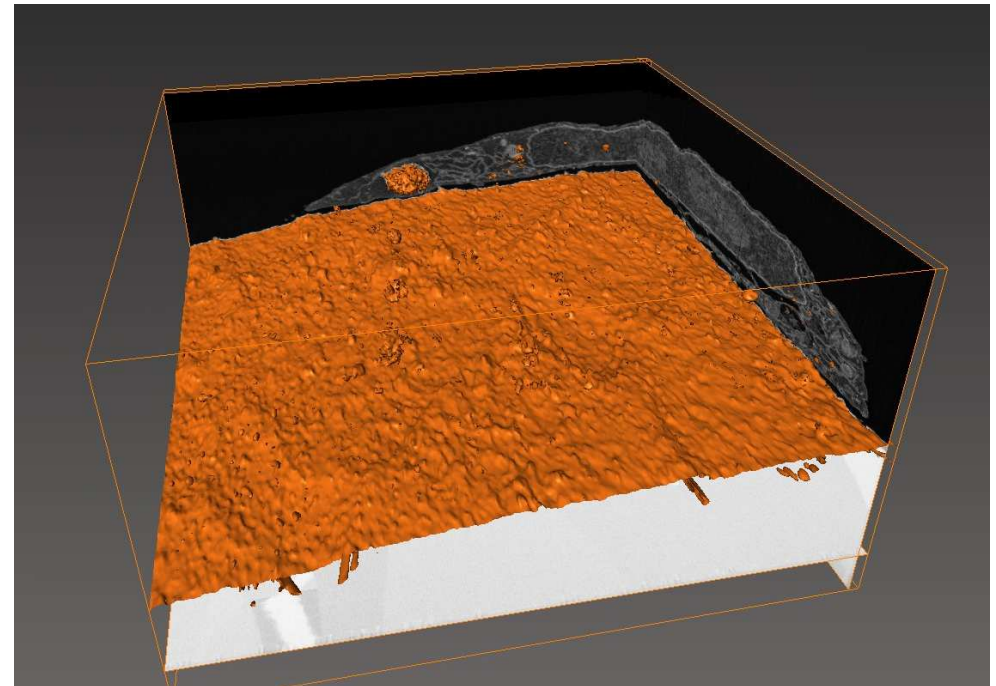


Segmentation based on grey levels

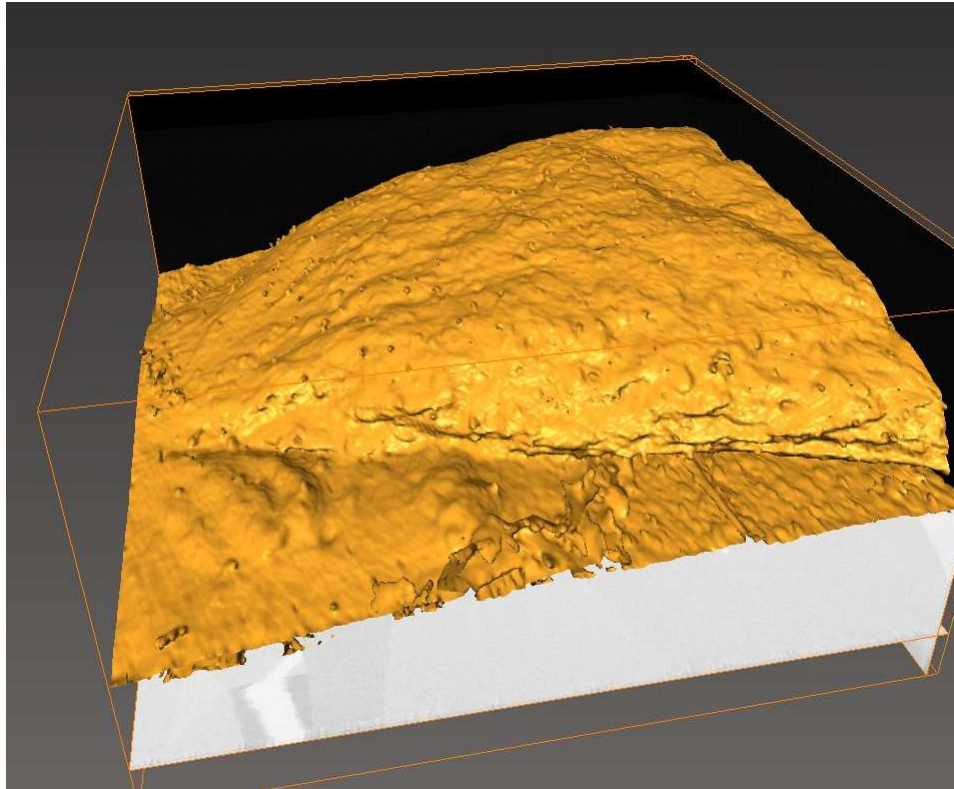
Medical steel



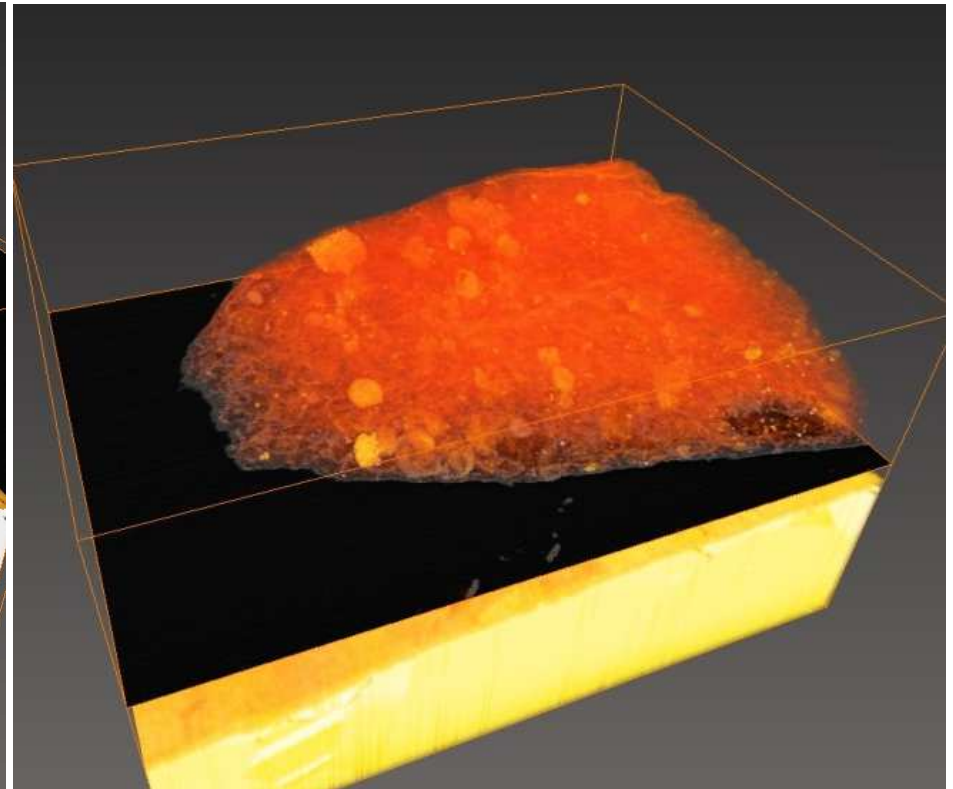
Ceramic coating: TiO_2

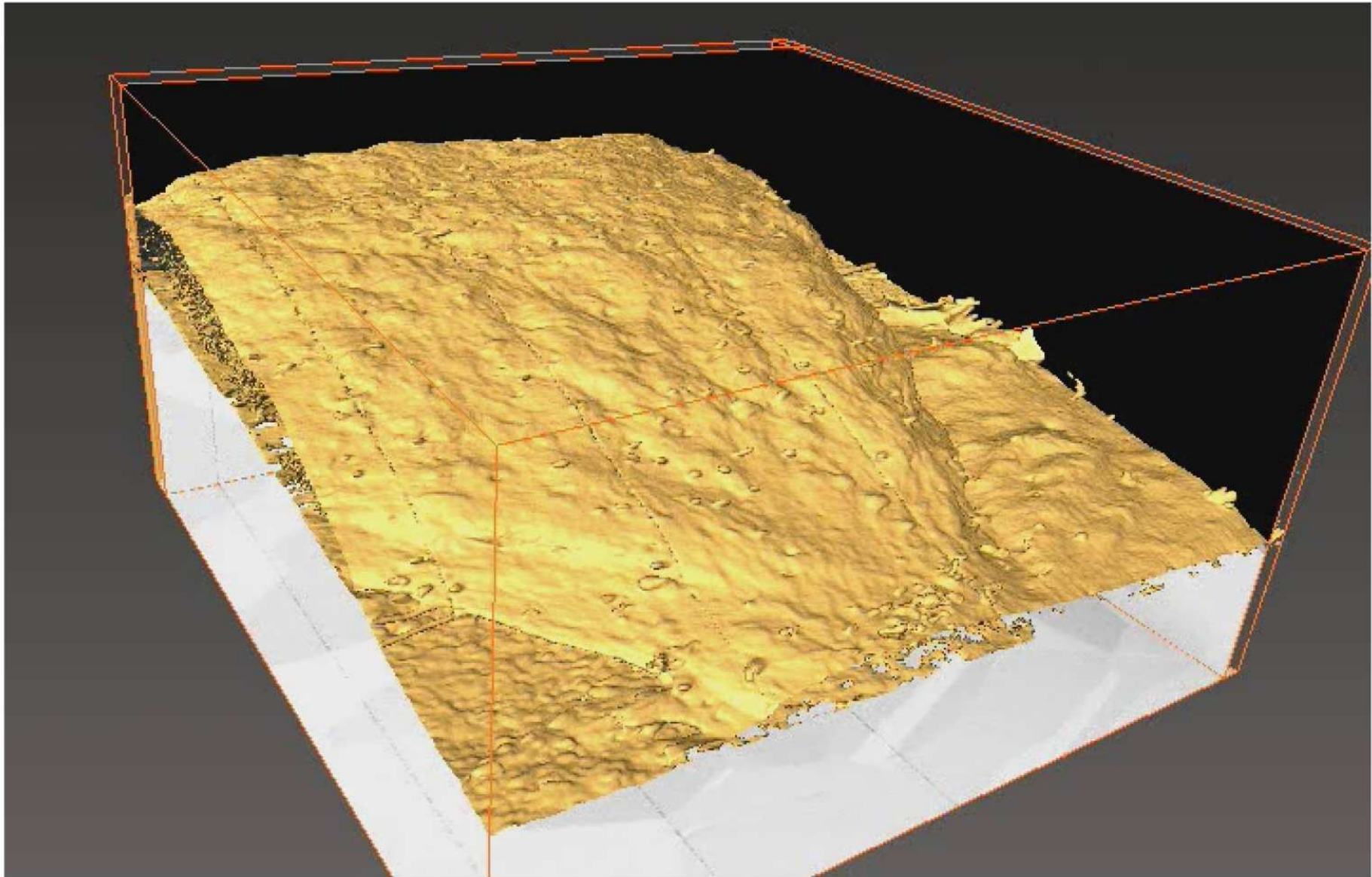


Cell outer membrane



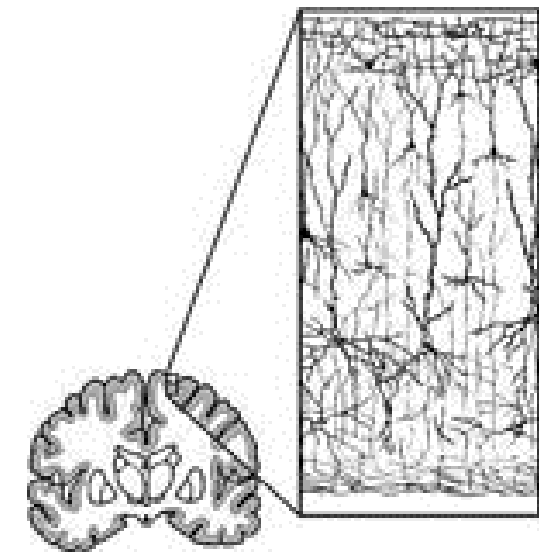
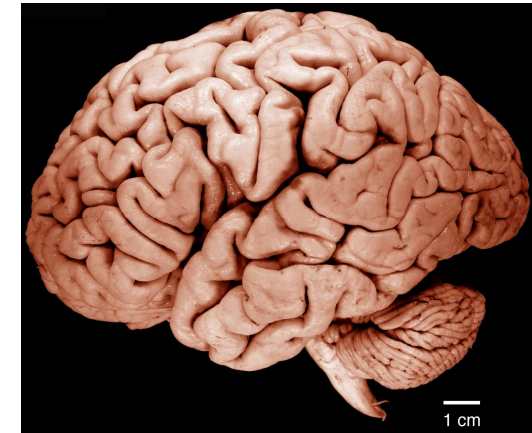
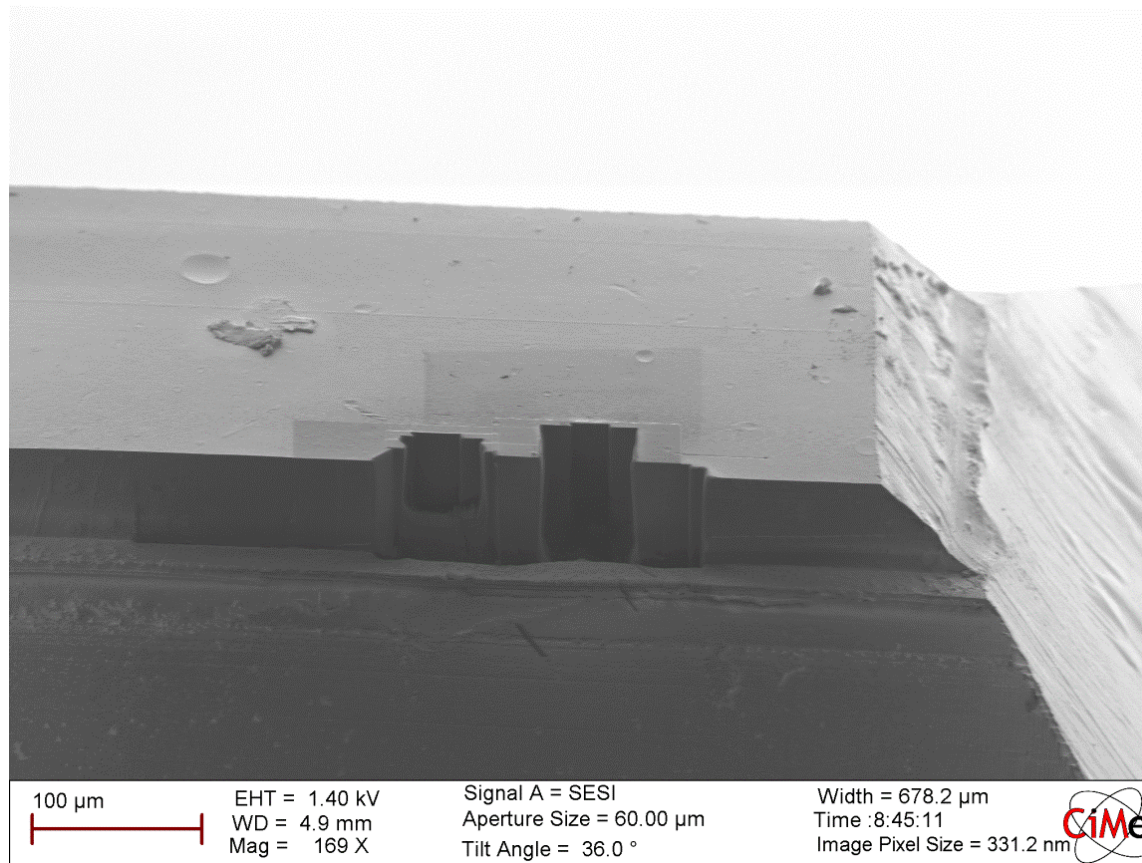
and more...





A little bit more complex....

brain tissue, resin embedded

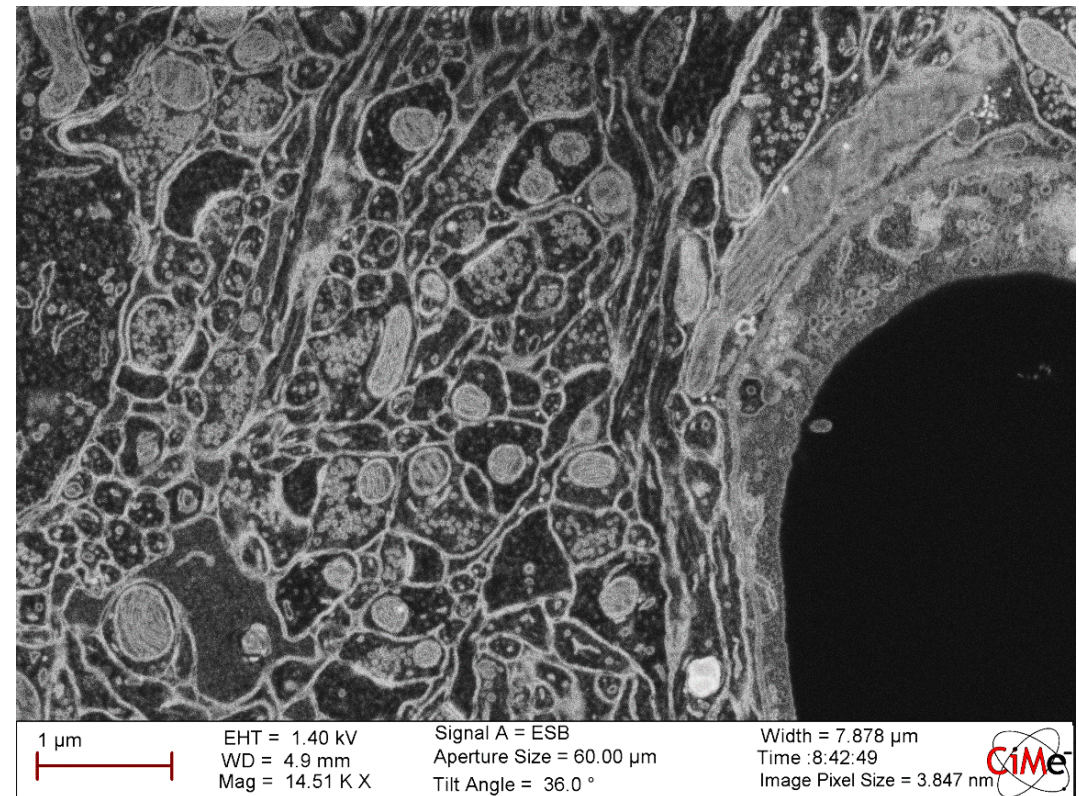
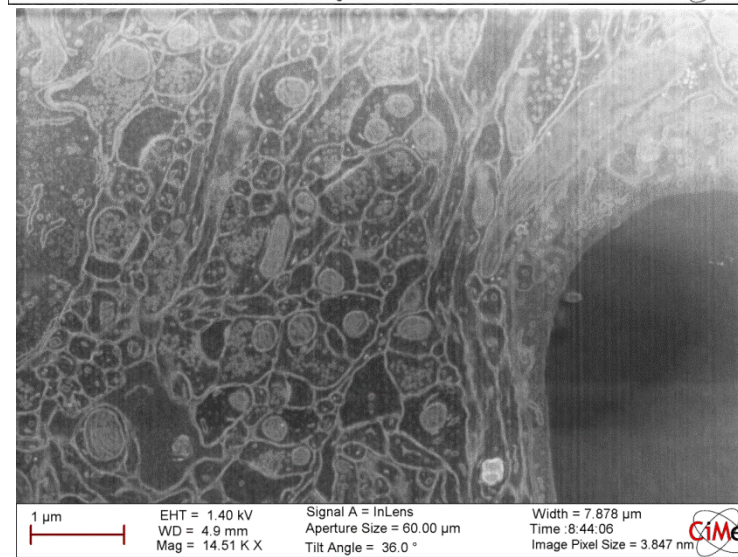
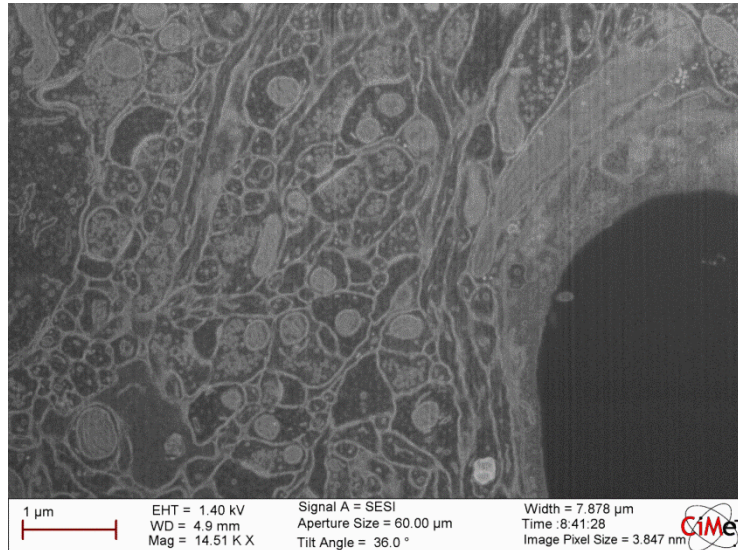


Which detector...?

In-chamber SE (Everhard-Thornley)

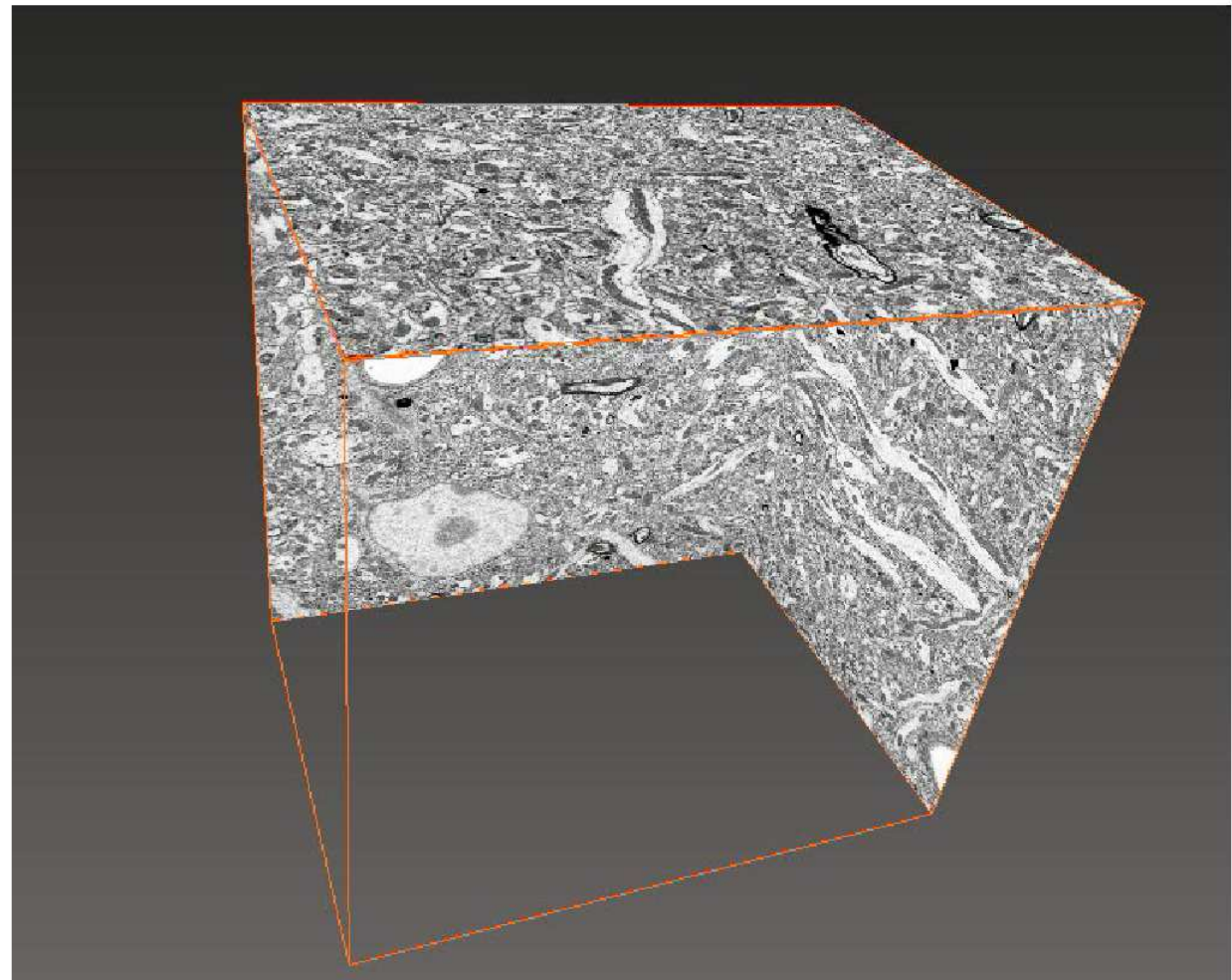
in-Lens SE

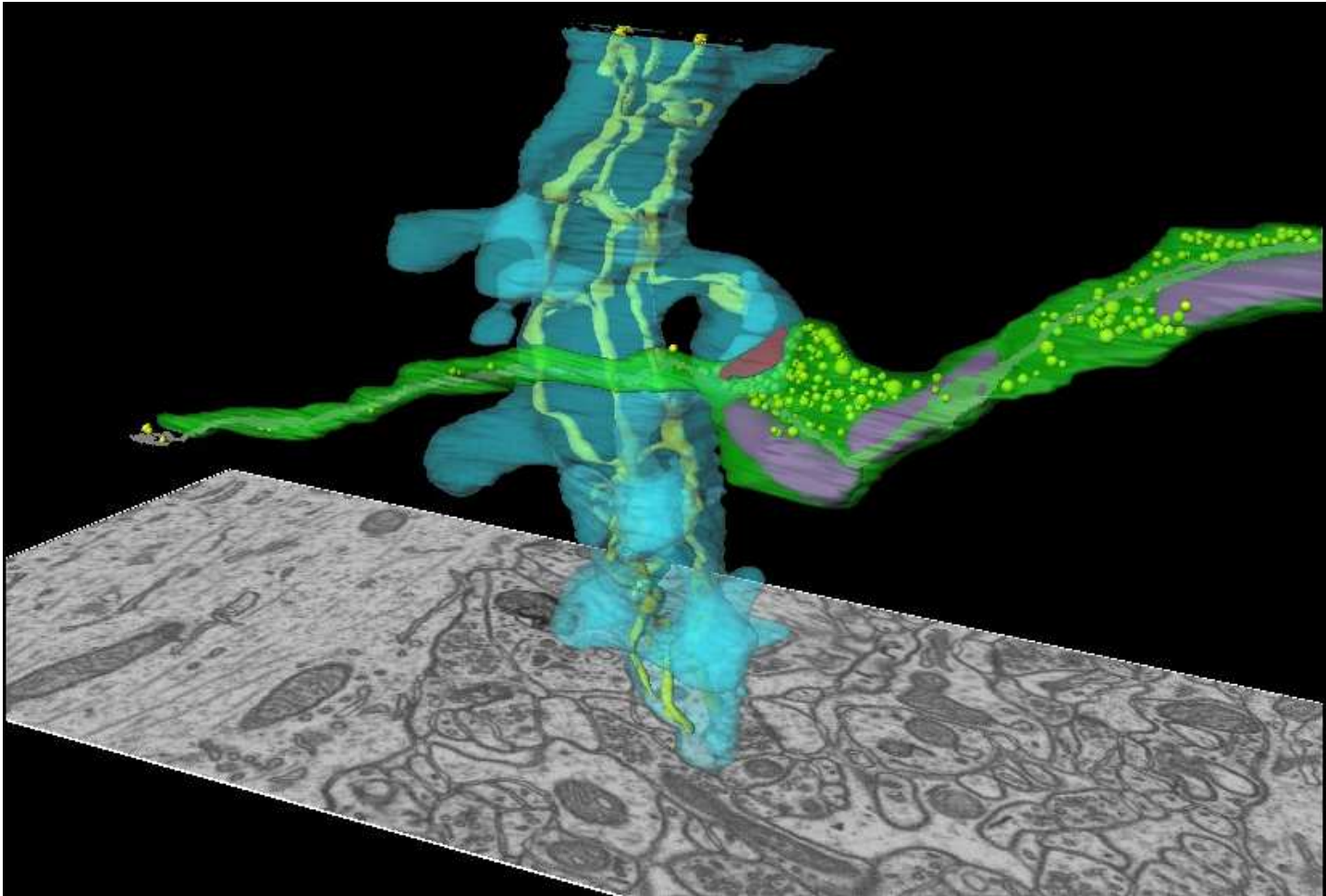
in-Lens BSE (energy selective)



Big volumes

- Voxel: 7.5x7.5x7.5nm
- Image 3096x2304
- 3300 slices (48hours)
- 23x17x24 um
- 9700um³
- ~7000 synapses
- 23Gb data



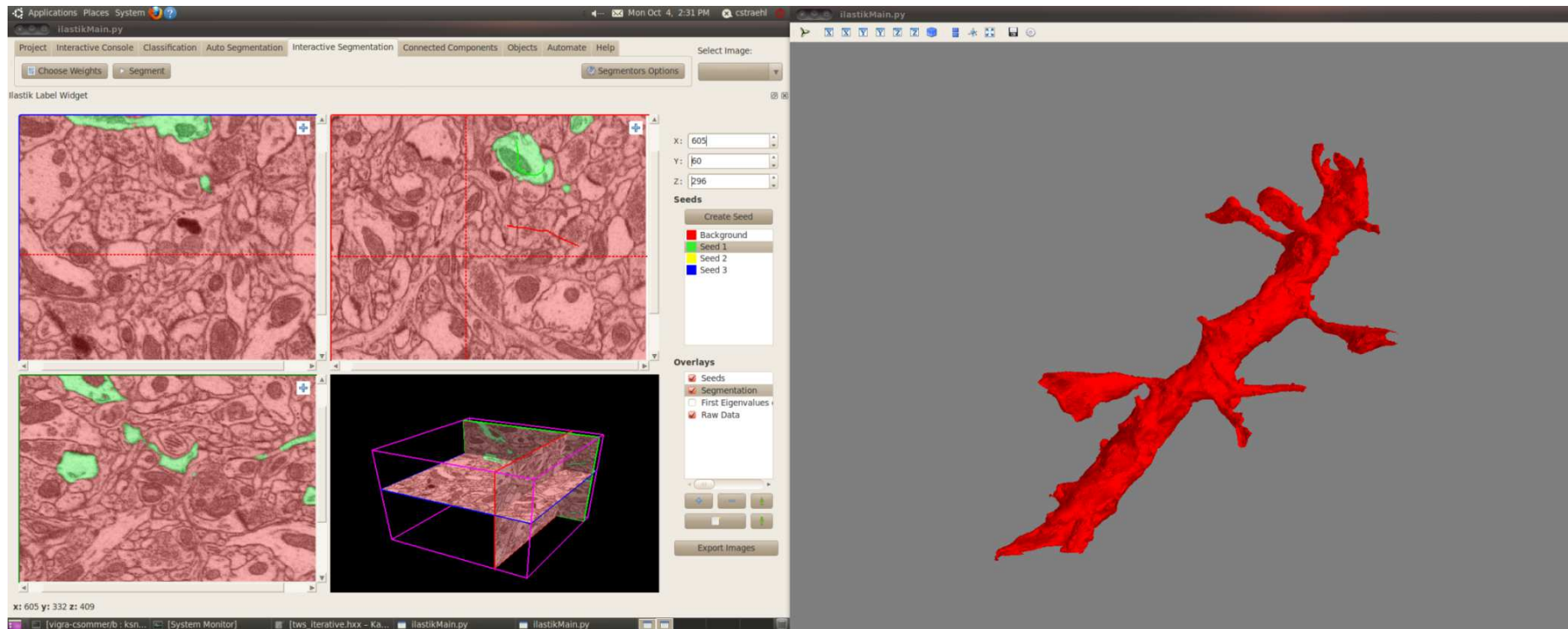


Reconstruction:
Christel Genoud

2weeks of work

Automated segmentation of neuronal structures

Ilastik v0.5 - Fred Hamprecht, University of Heidelberg



FIB Nanotomography in life science

- Specimen preparation (fixation, staining, dehydration, resin infiltration same as for BIO-TEM)
- Image contrast and resolution TEM quality
- Stable and reliable automated acquisition (less artifacts than ultra-microtomy)

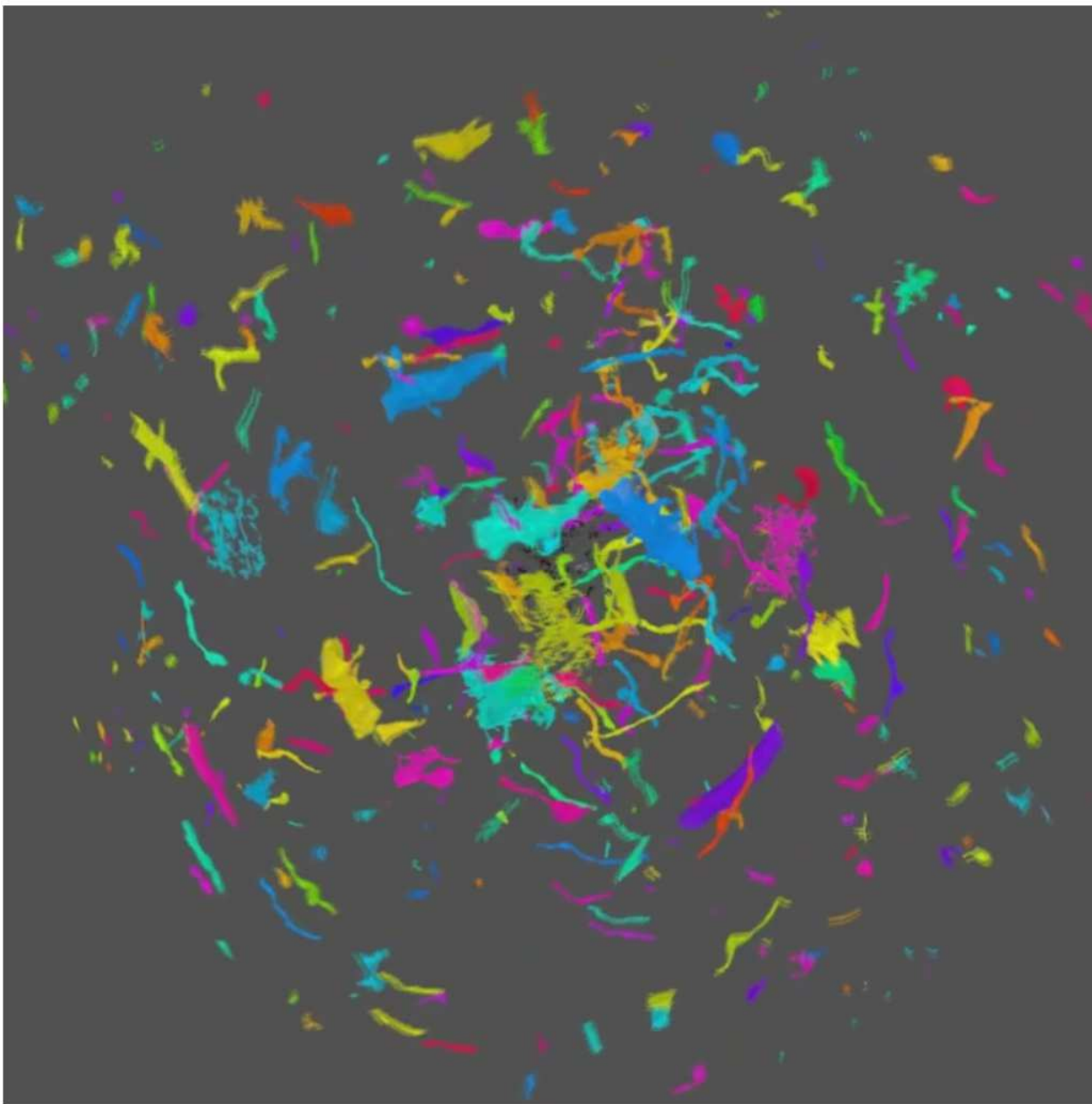


Image processing in 3D

- Start with a “good” image (low noise, equilibrated histogram, right detector)
- 2D processing (shading correction)
- Image alignment (registration)
- Cropping (subvolume)
- Noise reduction (2D or 3D)

- Segmentation (most important)
- Visualization (strong area for commercial software)